Histamine H₃ Receptor as a Drug Discovery Target

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Introduction

Ever since its pharmacologic discovery in 1983,¹ the histamine H₃ receptor has provided a tantalizing target for drug research, 25 years later still to be validated by an approved and marketed product. The H₃ receptor is one of the four currently known histamine receptor subtypes (H1-H4). Successful modulation of the histamine effects in the human body was initially achieved through the development of drugs acting at H₁ receptors, which were designed to mitigate the consequences of histamine release in an allergic reaction. The Claritin-Zyrtec-Allegra trio epitomizing the new generation of nonsedating antihistamines² went on to gain tremendous therapeutic and commercial success. At the same time a different class of drugs was shown to mediate secretion of gastric acid through selective antagonism of the H₂ receptors, a discovery that eventually led to the development of Tagamet and Zantac.³ The success of these drugs understandably heightened interest in the therapeutic and commercial potential of ligands of the H₃ and of the more recently discovered H₄ receptor.^{4,5} With pharmaceutical industry efforts naturally delayed in the early years by the challenges of H₃ biology, including lack of molecular identification of the human H₃ receptor, medicinal

chemistry programs finally shifted into high gear in the early 2000s. Importantly, the H₃ receptor position at the crossroads of neurotransmission suggests an abundance of therapeutic applications for small-molecule receptor modulators. Clearly a challenge to the tractability of in vitro and in vivo data, the complexity of H₃ biology has nevertheless set up expectations of a large payout at the end of the journey. While in mid-2009 this journey still remains to be successfully completed, a number of ongoing H₃ clinical programs, including several in phase II, an abundance of H₃ publications, and extensive H₃ patenting activity testify to the high level of interest in this area. Needless to say, the current medicinal chemistry programs are based on the monumental amount of research effort that has gone into establishing various aspects of H₃ receptor biology.

H₃ Receptor Pharmacology and Signaling Pathways

The initial role of the H₃ receptor was established by Arrang and co-workers to be that of a presynaptic autoreceptor mediating negative feedback of histamine release in rat brain.¹ Fifteen years later in 1999, Lovenberg and co-workers identified a GPCR^{*a*} clone similar to biogenic amine receptors through a homology search of expressed sequence tag databases.⁶ This clone revealed a pharmacological profile corresponding to that previously established for the H₃ receptor.⁷ The availability of human H₃ cDNA facilitated further research into the structural makeup and molecular signaling mechanisms of the H₃ receptor. The H₃ receptor shows little similarity to its predecessors (20% homology to H₁ and H₂) with more similarity (60% homology) to the recently discovered H₄ receptor, though with different expression pattern in vivo.

While the H₃ receptor is a classic GPCR with seven transmembrane domains, its isoform signature is intensely complex. This aspect of H₃ pharmacology has been reviewed on more than one occasion by Leurs and co-workers.^{8,9} While many human H₃ receptor isoforms have been identified, the full length 445 configuration appears to be the most functionally dominant and abundantly expressed. In addition, at least another 19 isoforms, a number of them functional as well, have been reported for the human receptor. Several additional isoforms have also been identified in rat, guinea pig, and mouse. Not surprisingly, complex isoform composition and their different expression patterns are often invoked in connection with unexpected or inconsistent in vivo H₃ biology. However, at this point there is a clear need for more data to substantiate these correlations or perhaps to provide a simpler explanation. While selective isoform targeting could in theory contribute to the selectivity of H₃ ligands, in practice

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^{*a*} Abbreviations: ^{-/-}, gene knockout; 6-OHDA, 6-hydroxydopamine; AD, Alzheimer's disease; ADHD, attention-deficit hyperactivity disorder; Akt, serine/threonine protein kinase; ALDH, aldehyde dehydrogenase; ATP, adenosine triphosphate; BA, bioavailability; BAT, brown adipose tissue; b.i.d., twice a day; BMI, body mass index; B/P, brain/ plasma; cAMP, cyclic adenosine monophosphate; CNS, central nervous system; COX, cyclooxygenase; CSF, cerebrospinal fluid; CYP450, cytochrome P450; DIO, diet-induced obesity; DREM, direct rapid eye movement; EDS, excessive daytime sleepiness; ERK, extracellular signalrelated kinase; FDA, Food and Drug Administration; GABA, γ-aminobutyric acid; GPCR, G-protein-coupled receptor; GSK-3 β , glycogen synthase kinase- 3β ; GTP γ S, guanosine 5'-O-(γ -thio)triphosphate; hDAT, human dopamine transporter; HDC, histidine decarboxylase; hERG, human ether-a-go-go-related gene; HNMT, histamine *N*-methyltransferase; hH_3R , human H_3 receptor; hNET, human norepinephrine transporter; hSERT, human serotonin transporter; HSV, herpes simple virus; icv, intracerebroventricular; IgE, immunoglobulin E; IL, interleukin; ip, intraperitoneal; iv, intravenous; L-dopa, 3,4-dihydroxyphenyl-L-alanine; MAO B, monoamine oxidase B; MED, minimal efficacious dose; mRNA, messenger ribonucleic acid; MS, metabolic syndrome; NHE, Na⁺/H⁺ exchanger; NIDCD, National Institute on Deafness and Other Communication Disorders; NK, neurokinin; NMDA, N-methyl-D-aspartic acid; NREM, nonrapid eye movement; PD, Parkinson's disease; PET, positron emission tomography; PGC1, proliferator-activated receptor- γ coactivator 1; PI3K, phosphatidylinositol 3-kinase; PK, pharmacokinetic; PLA₂, phospholipase A₂; po, oral; q.d., once a day; RAMH, R-αmethylhistamine; REM, rapid eye movement; rH3R, rat H3 receptor; sc, subcutaneous; SAR, structure-activity relationship; SSRI, selective serotonin reuptake inhibitor; $t_{1/2}$, half-life; TM, transmembrane domain; WT, wild-type.



Figure 1. Biosynthesis and metabolism of histamine: HDC, histidine decarboxylase; HNMT, histamine *N*-methyltransferase; MAO B, monoamine oxidase B; ALDH, aldehyde dehydrogenase.



Figure 2. Biological role of H₃ receptors: HA, histamine; ACh, acetylcholine; HDC, histidine decarboxylase; NE, norepinephrine.

it remains just a vague possibility, especially in view of the questionable functional significance of many isoforms. Additionally, several instances of genetic polymorphism for the human H_3 receptor have been reported.^{6,10}

Studies into H₃ receptor expression patterns have revealed its abundant presence in the CNS of rodents and humans, lower expression in the sympathetic nervous system, and yet lower expression in peripheral tissues. Detection of H₃ receptor mRNA through in situ hybridization has added to the insights into receptor distribution, earlier approached using autoradiography.^{11,12} The H₃ receptor is largely expressed on, although not totally confined to, histaminergic neurons, the site of histamine synthesis in animals and humans (Figure 1). Histamine is formed in vivo from L-histidine by histidine decarboxylase (HDC) and released in the neuronal synapse, then deactivated by metabolic conversion to tele-methylhistamine by histamine N-methyltransferase (HNMT). Further metabolic oxidation by monoamine oxidase B (MAO B) and aldehyde dehydrogenase (ALDH) leads to tele-methylimidazoleacetic acid.¹³ In the CNS of humans and rodents, the histaminergic neurons originate in the tuberomamillary

nucleus of the posterior hypothalamus, from which extensive projections reach into all major areas of the brain and parts of the spinal cord.^{14,15} As a result, the H₃ receptors are present in cerebral cortex, hippocampus, amygdala, striatum, and basal ganglia, in addition to hypothalamus. The H₃ receptor role appears to be mediation of functional effects of histamine in different tissues. As a neuronal presynaptic autoreceptor, H₃ provides negative feedback regulation of histamine release in the neuronal synapse, which moderates histamine inhibitory control of the firing of histamine neurons,¹⁶ as well as its actions at the postsynaptic histamine receptors (vide infra). CNS histamine as a neurotransmitter has been implicated in diverse responses, including arousal, cognition, fluid, food and temperature homeostasis, cardiovascular control, anxiety, and pain perception, inexorably linking the central H₃ receptor system to these fundamental physiological processes.¹⁷ In addition to a direct role in histamine signaling, the H₃ receptor is also involved in regulation of other neurotransmitter systems directly as a presynaptic heteroreceptor (Figure 2) or indirectly by affecting levels of histamine, which then acts on postsynaptic receptors. H₃ heteroreceptor modulation has the potential



Figure 3. Early synthetic H₃ ligands.

to affect classic biogenic amine neurotransmitters such as glutamate,¹⁸ acetylcholine,¹⁹ norepinephrine,²⁰ dopamine,²¹ GABA,²² and serotonin,²³ conceivably setting up a variety of therapeutic applications for the H₃ receptor. Centrally or peripherally, the effects of H₃ activation or blockade, as evidenced by H₃ biology and discussed below, often seem to be dependent on the exact nature of the tissue, as defined by locale, cell type, and species. In addition to acting as a neurotransmitter in the sympathetic nervous system, histamine in the periphery has been linked to mediation of inflammatory, immunosuppressive, and gastrointestinal effects. Outside the neuronal system, significant sources of histamine are enterochromaffin-like cells in the gut, where its release upon gastral and vagal simulation promotes acid secretion, and mast cells and basophils, where histamine release, triggered by allergens, and accompanying downstream effects constitute inflammatory response during an allergic reaction (vide infra). However, immune and inflammatory responses to histamine in leukocytes and lymphocytes are mediated by H₁, H₂, and H₄ receptors, as H₃ receptors appear not to be expressed on these cells.⁵ Histamine action on mast cells has been suggested to cause chemotaxis, but not degranulation, again mediated by H_4 rather than H_3 receptors, which appear not to be expressed on murine²⁴ or human mast cells,²⁵ although H₃ receptors have been implicated in morphological changes and histamine release from rat brain mast cells.²⁶ Also suspect is the role of H₃ in human gastrointestinal tissues (vide infra). Separately, the H₃ receptor may have a potentially important, even if somewhat indirect, role in the mediation of effects of allergic rhinitis through regulation of norepinephrine levels in the sympathetic nervous system and, consequently, vascular tone in nasal mucosa linked to nasal congestion.27

Early synthetic H₃ agonists, used for receptor characterization, included very close analogues of histamine itself, such as *R*- α -methylhistamine 1 (RAMH) and *N*-methylhistamine 2, as well as more elaborate examples imetit 3 and immepip 4 (Figure 3).^{28,29} All of these compounds have shown some activity at other histamine as well as other biogenic amine receptors, which along with low oral bioavailability and rapid metabolism, has limited their utilization in vivo (as an example, see discussion on clinical trials for migraine with 1, vide infra). However, they have figured prominently in historic in vitro pharmacological studies. Similar to archetypal agonists, all the early H₃ antagonists were based on the 4-substituted imidazole motif. Most prominent among them are thioperamide 5, clobenpropit 6, and ciproxifan 7 (Figure 3).^{28,30,31} Proxyfan 8, initially identified as a neutral H₃ antagonist, has been recharacterized as a protean agonist, its functional profile dependent on the assay system used.³² While histamine and H_3 agonists have been generally linked to the down-regulation of histamine and other neurotransmitter release in tissue preparations and in vivo and while antagonists have generally had the opposite pharmacologic response, the underlying complexity of H₃ receptor biology has often rendered exact agonist-antagonist classification challenging and dependent on the particular assay. In addition, most antagonists were reclassified as inverse agonists in view of their effect on H₃R constitutive activity in some, but not necessarily all, functional assays.^{33,34} At the same time, the rate of histamine turnover appears to be another differentiating factor: concomitant to increasing histamine levels, some (e.g., 5) but apparently not all H₃ antagonists also increase levels of telemethylhistamine.^{35,36} Although judged a poor correlate of the antagonist activity, this tele-methylhistamine-elevating effect is observed with the most recent H₃ antagonists (vide infra).

On the molecular level the H_3 receptor is associated with several major pathways of signal transduction. These pathways have been repeatedly reviewed in the literature (in particular, recently in much detail by Leurs and co-workers)³⁷ and are also discussed in the Supporting Information and illustrated in Figure 4 of this manuscript. Connections between the biochemical signaling mechanisms and in vivo effects of the H_3 agonists and antagonists, whether hypothetical or clearly supported by data, are contemplated throughout the review.

Regulation of neurotransmitter release is considered the main underlying cause of the observed and/or expected therapeutic effects of H₃ receptor ligands in animal models or in humans. Changes in histamine levels result in variable activation of postsynaptic histamine receptors with key therapeutically relevant effects mediated by H1 but with some roles also reserved for H_2 and, possibly, postsynaptic H_3 receptors³⁸ (see Figure 2). The connection between H₃ and H₁ signaling has been demonstrated through the dependence of typical H₃-mediated effects on the presence of functional H₁ receptors in the H₁ knockout $(H_1^{-/-})$ vs wild-type mouse experiments.³⁹ At the same time, H₃ heteroreceptors found on nonhistaminergic neurons regulate other neurotransmitters. Increases in the levels of neurotransmitters in the presence of an H₃ antagonist are generally rationalized by the antagonist's blockade of histamine's negative feedback via H₃ (resulting in the continued neurotransmitter release, histamine itself included) and supplemented by



Figure 4. Molecular signaling pathways of the H3 receptor: AC, adenylyl cyclase; Akt, serine/threonine protein kinase; CaMKII, calcium/ calmodulin-dependent protein kinase type II; cAMP, cyclic adenosine 3',5'-monophosphate; CREB, cAMP responsive element-binding protein; GSK- 3β , glycogen synthase kinase-3b; ERK, extracellular signal-related kinase; HDC, histidine decarboxylase; JNK, Jun N-terminal kinase; MAPK, mitogen-activated protein kinase; PI3K, phosphoinositide 3-kinase; PKA, protein kinase A; PLA2, phospholipase A2.

histamine activation of H₁ receptors, with the latter sometimes the major factor. Thus, for example, an increase in the levels of acetylcholine was reported upon rat brain superfusion with histamine, despite the envisioned histamine's H₃ agonist action on cholinergic neurons. This acetylcholine-elevating effect was blocked by an H1 antagonist but not affected by H2 or H3 antagonists, suggesting the overriding role of H1 receptors, at least in this experiment.^{19,40,41} However, the relationship between H₃ receptor activation/inhibition and the resulting neurotransmitter levels is often complex and multitiered. In addition to direct modulation by H₃ heteroreceptors on appropriate neurons, indirect pathways via other neurotransmitters have been suggested. It was shown, for example, that inhibition of acetylcholine release from rat cortex by H₃ agonists⁴² may be GABAergic in nature. The effect of 4 was reversed by a GABA antagonist, suggesting positive coupling of H₃ heteroreceptors to GABA, which presumably in turn inhibits release of acetylcholine.⁴³ At the same time, the H_3 receptor effect does not appear to be uniform throughout the brain. Thus, several studies suggest the opposite effect in basolateral amygdala, with H₃ agonists enhancing and H_3 antagonists reducing acetylcholine release.^{44,45} The complex nature of interactions between different neurotransmitter systems is well illustrated by the effects observed in H_3 knockout (^{-/-}) mice. Not surprisingly, histamine signaling is altered because of its reduced levels and reduced expression of H₁ receptors reported in a study of energy homeostasis effects in $H_3^{-/-}$ mice. A substantial concomitant increase in tele-methylhistamine levels (up to 135% in the hypothalamus) was also noted.⁴⁶ In a different study, which addressed behavioral changes in $H_3^{-/-}$ mice, the not easily explainable 42% reduction in brain cortex histamine content was accompanied by a reduced stimulatory effect of methamphetamine on locomotor activity, implicating decreased dopaminergic and/or

serotonergic activity relative to $H_3^{+/+}$ animals. In this case, the density of H₁ receptors was reported to be unaffected by the gene knockout.⁴⁷ $H_3^{-/-}$ mice also showed resistance to the amnesic effects of the cholinergic antagonist scopolamine, suggesting perturbed cholinergic signaling as well. While these signaling interconnections appear to be extensive, more data are clearly needed to establish their exact nature. In addition to the receptor cross-talk at the functional level (concomitant response from receptors, different from the one directly activated, and arising from signaling molecules being shared across different signaling pathways), direct intramembrane interactions between different aminergic GPCRs at the molecular level, resulting in the formation of heteromeric complexes, have been reported.⁴⁸ This scenario was invoked in one particular example, where H₃ receptor displayed an antagonistic interaction with the D_2 receptor, and selective H₃ agonist 1 weakened D₂ receptor binding of the selective D₂ agonist quinpirole in sheep striatal membranes in vitro.⁴⁹ Moreover, the H₃ selective agonist 3 inhibited locomotor activation induced in reserpinized mice by the selective D_2 agonist quinpirole, while the H_3 selective antagonist 5 potentiated it. Such interactions (direct intramembrane or functional) may be of interest to drug discovery in view of the interconnecting nature and important roles of both histaminergic and dopaminergic neurotransmission in memory, attention, and psychiatric disorders (vide infra).⁵⁰

While the abundance of cellular signaling pathways potentially influenced by H_3 receptor-ligand binding interactions would provide rationale for a variety of in vivo directions, the nature of H_3 biology often gives rise to complex in vivo profiles. This wide spectrum of in vivo effects (both already discovered and still targeted) should also be appreciated in context with the diversity of structural types developed over the 15-plus years of industrywide medicinal chemistry efforts.



Figure 5. Historic H₃ agonists based on 4-substituted imidazole.



Figure 6. Historic H₃ antagonists/inverse agonists and protean ligands based on 4-substituted imidazole.

Historic H₃ Ligands

Over the past 20 years, numerous imidazole-based H₃ ligands have been described. Still in line with the archetypal 4-substituted imidazole scaffold, agonists and antagonists, in addition to those already shown in Figure 3 and used extensively in the receptor characterization, have been identified (Figures 5 and 6).^{8,51} As already discussed, reclassification of many H₃ antagonists as inverse agonists, in particular the most commonly used compounds 5-7, followed studies on H₃R constitutive activity. Cipralisant 16 (GT-2331, Perceptin, vide infra), designated a protean agonist similar to 8, was one of the first H₃ compounds to advance to the clinic, reaching phase II enrollment for ADHD before progress was halted in 2002.⁵² The imidazole-based natural product verongamine **21** displayed moderate H₃ binding in guinea pig membranes and was used as a template for further discoveries.⁵³ Numerous reviews have been published summarizing early work in the field as well as issues relating to selectivity versus H4, differentiated receptor binding in various species (rat vs human), off-target affinities, and P450 inhibition caused by the imidaz-ole ring. $^{8,51,53-57}$

Therapeutic Applications of H₃ Agonists

A variety of potential therapeutic applications for H_3 agonists have been reported including inflammation, migraine, asthma, insomnia, pain, ulcer, hypertension, immunological regulation, and ischemic arrhythmias.^{8,52,58} Generally associated with negative neurotransmitter modulation, H_3 agonists

are expected to provide beneficial effects in cases where elevated neurotransmitter levels may be the reason for pathological conditions. In that regard, histamine's role in the maintenance of the awake state, nociceptive signaling, and inflammatory mediation sets it up as a target of therapeutic interference. For example, superfusion of rat basal forebrain with histamine produces a significant dose-dependent increase in wakefulness, presumably through excitation of cholinergic and noncholinergic neurons.⁵⁹ This arousal action, mediated by postsynaptic H_1 receptors and illustrated by the sedative effects of brain penetrating H_1 antagonists,^{60,61} has been associated on its extreme end with insomnia.⁶⁴ Compounds **3**, **13** (BP294, also FUB94), and 14 (Sch 50971) have shown increased sleep time and/or sedation in preclinical animal models, effects that were neutralized by H_3 antagonists.^{63–65} However, a recent sleep study with 4 showed little sleep promoting effect in rats despite a significant decrease in brain histamine concentrations.⁶⁶ At the same time, insomnia has been mentioned as the major adverse event in a clinical trial with the H₃ antagonist 59 (PF-03654746) (vide infra).

Histamine's involvement in neurogenic inflammation in various tissues, characterized by afferent nerve-mediated vasodilation, increased vascular permeability, plasma leakage, smooth muscle contraction, increased inflammatory cytokine production, and immune cell recruitment,⁵ implicates it in mediation of pathological processes associated with allergic inflammation,⁶⁷ asthma,⁶⁸ and migraine.⁶⁹ As discussed above, relevance of the H₃ receptor agonists is in presynaptic regulation of histamine, the postsynaptic inflammatory effects of which appear to be

mediated via H1 and H4 receptors. Thus, intradermal administration of histamine induced scratching behavior in mice, reduced by H_1 and H_4 , but not H_2 antagonists.⁷⁰ While the effect of H_3 antagonist $-H_4$ agonist **6** in this study was attributed to its H_4 agonist activity, other studies^{71,72} have suggested that **6** causes pruritis and allergic rhinitis (inhibited by NK1 receptor antagonists) through H₃ antagonist-induced release of neuropeptide substance P.⁷³ In general, the effects of H₃ receptor modulation may extend, directly or indirectly via histamine, to other neurotransmitters and mediators, critical or contributing to the pathology of neurogenic inflammation. Thus, in addition to histamine and substance P, other H₃-modulated neurotransmitters (e.g., acetylcholine, serotonin) also cause bronchoconstriction in allergic responses by effecting smooth muscle contraction. Alleviation of this condition by H₃ agonists (due to their moderating effect on neurotransmitter release) may be expected; however, because of multiple possibilities, the exact mechanism of action may be difficult to establish. For example, the H₃ agonist 1 acted as a smooth muscle relaxant in the perfused guinea pig bronchioles and lung strips under resting tone,⁷⁴ the effect presumably arising from the inhibition of acetylcholine and histamine background release. However, 1 also reversed bronchoconstriction induced by electrical field stimulation under the conditions of postsynaptic histamine and muscarinic receptor blockade by their respective antagonists,75 leaving open the interpretation of the exact mechanism. In addition, 1 reversed bronchoconstriction caused by exogenously administered histamine or acetylcholine,76 the effect hypothesized to result from increased levels of metabolites of arachidonic acid on the basis of dose-dependent bronchoconstriction increase by COX inhibitor indomethacin.⁷⁷ The bronchodilatory effect of 1 in all cases was blocked by the H_3 antagonist 5. In another study, 1 and 3 reduced endothelial permeability in acetylcholine-induced pulmonary edema in isolated perfused rabbit lungs, a finding attributed to the H₃ agonist effect on substance P levels.⁷⁸ Compound 14 alleviated sneezing and nasal rubbing in a mouse model of allergic rhinitis⁷² and was reported active in a central neurogenic vascular inflammatory model of migraine in the guinea pig.⁶⁵ As a potential drug candidate, 1 displays good in vitro potency, but its poor human oral absorption, poor brain penetration, and rapid metabolism by HNMT were discovered in early clinical trials.^{54,79,80} A prodrug of 1, benzhydrylimine 13, displayed improved human pharmacokinetic properties over the parent compound (plasma half-life $t_{1/2} > 24$ h)⁷⁹ and reached phase II clinical trials for asthma and migraine; however, lack of information on compound progression since 1996 moderates enthusiasm for these indications.⁵² Elsewhere, in a phase II trial featuring unusually low doses (1-3 ng twice weekly, subcutaneous (sc)), the nonselective H_3-H_1 agonist 2^1 decreased the frequency, intensity, and duration of migraine attacks. However, intense headaches were reported at higher doses (4 ng in migraine patients and 10 ng, previously, in phase I healthy volunteers), possibly because of neurogenic edema caused by H₁ agonism. Although phase III results, disclosed in 2006, apparently confirmed antimigraine activity of *N*-methylhistamine without evidence of adverse effects, ^{58,82} there seems to have been no filing for registration.

While the role of H₃ receptors in nociception has been investigated, the resulting evidence is conflicting. Sensitivity to noxious stimuli was greatly reduced in $H_1^{-/-83}$ and histidine decarboxylase knockout (HDC^{-/-}) mice,⁸⁴ while administration of exogenous histamine in the latter case significantly induced nociceptive behavior, inhibited, in turn, by the H₁ antagonist pyrilamine. Antinociceptive activity has been reported with

 H_3 agonists 13^{79} and $4^{85,86}$ in rats, although the effects were not general (model- and species-specific) and not completely understood. However, these data seem to be contradicted by the fact of analgesic effects observed with histamine itself, histidine, the HNMT inhibitor metoprine as well as H_3 antagonists.^{87,88} In general the outcome may certainly be affected by the physiological differences between acute nociceptive and chronic inflammatory and neuropathic pain,⁸⁹ the latter being the ostensible clinical target of H_3 antagonists (vide infra).

Modulation of norepinephrine release in the cardiovascular system by H₃ receptors located on sympathetic nerves in heart and blood vessels suggests the use of H₃ agonists for the treatment of cardiac dysfunction and ventricular arrhythmias associated with myocardial ischemia and triggered by excessive norepinephrine levels.⁹⁰ As discussed above, activation of H_3 receptors reduces norepinephrine release by inhibiting both Ca^{2+} -mediated exocytotic⁹¹ and carrier-mediated (Na⁺/H⁺ exchanger (NHE) activated) mechanisms,⁹² the first being associated with acute and the second with protracted myocardial ischemic condition. On the basis of isolated guinea pig and human heart tissue experiments, the role of 3 in the reduction of norepinephrine release was established for the protracted but not the acute condition, where H₃ receptors already appear to be fully activated.⁹⁰ In addition, the role of H₃ agonists in mediating cardiovascular effects extends to the presynaptic regulation of histamine release in the heart. Positive chronotropic and inotropic effects of histamine (increase in the rate and force of heart muscle contraction) in dogs and humans have been linked to its action on postsynaptic (H₁, H₂) receptors.⁹³ In that regard, use of H₃ agonists under the conditions of cardiovascular distress could inhibit release of histamine and reduce its cardiac stimulatory and arrythmogenic effects. Despite all these data or perhaps precisely because of histamine's potentially fundamental role in maintaining the status quo, there have been no attempts so far to develop an H₃ agonist for cardiovascular indications.

In addition to the research directions outlined above, an apparent connection between elevated histamine levels (presumably achieved as a result of exogenous histidine administration) and increased anxiety in mice (attenuated by an H_1 antagonist) has been reported in the elevated plus-maze test.⁹⁴ Conceivably, this would suggest a role for H_3 agonists as antianxiety agents, although no relevant data have been disclosed.

Therapeutic Applications of H₃ Antagonists/Inverse Agonists

A variety of potential therapeutic applications for H_3 antagonists/inverse agonists have been proposed including CNS disorders such as Alzheimer's disease (AD), schizophrenia, dementia, anxiety, tremor (Parkinson's disease (PD)), attention deficit hyperactivity disorder (ADHD), depression, mood disorders, sleep disorders (narcolepsy), pain/itch, stroke, epilepsy, as well as metabolic syndrome (MS, including obesity and diabetes), nasal inflammation (allergic rhinitis, congestion, allergy), and cancer.^{8,51,99,53,58,62,95–100}

In contrast to the early work in the field, most chemical series of current interest appear to be nonimidazole in nature, reflecting major disadvantages of the 4-substituted imidazole motif, including poor brain penetration and issues related to CYP450 inhibition, such as drug-drug interactions, extrapyramidal symptoms, liver toxicity, and inhibition of adrenal steroid synthesis.^{51,57} In addition, nonimidazoles tend to be



Figure 7. Miscellaneous non-H₃ compounds.





Figure 8. Early non-imidazole H₃ antagonists.

more selective for H₃ vs H₁, H₂, and H₄.¹⁰¹ In 1994, a weak nonimidazole H₃ antagonist, the marine natural product aplysamine-1 (23, Figure 8), was patented by Harbor Branch Oceanographic Institution.⁵¹ The compound contained a phenoxyalkyl-N,N-dialkylamine fragment that would later become an important reoccurring motif in the field. Starting in 1998, Ganellin and co-workers (University College, London) described a systematic SAR exploration that resulted in a series of compounds including 24 (UCL-1972), which showed good potency and a favorable pharmacokinetic profile.8,51,57 The academic FUB-UCL-INSERM-Bioproject consortium has provided many additional examples of potent nonimidazole H₃ antagonists including the potent analogues of 7, such as compounds 25 (UCL-2190) and 26 (UCL-2173). further confirming the emerging nonimidazole pharmacophore 27 as an exciting new area for H_3R research.⁵¹ Early work by Menge and co-workers (Vrije Universiteit) was based on the nootropic agent sabeluzole **28** (Figure 7) and led to the potent ligand **29** (VUF5391).^{8,51} The James Black Foundation also disclosed examples of early nonimidazole H₃ antagonists, including the selective and potent guanidinebased **30** (JB-98064).⁵¹ Modeled after the acetylcholinesterase inhibitor tacrine **31** (Figure 7), compound **32** (FUB-833) demonstrated dual H₃ antagonist–acetylcholinesterase inhibitory activity,¹⁰² whereas the truncated analogue **33** (FUB-836) was reported as the first in a new class of dual H₃ antagonist– HNMT inhibitors.¹⁰³ More details on early nonimidazole antagonists can be found in recent reviews.^{8,51,57}

Evolution of the prototypic H_3 antagonist pharmacophore has been discussed elsewhere.⁵¹ The current generally accepted model incorporates a basic amine motif separated by several atoms from the central, typically hydrophobic, core, which is joined on the other side by a structurally variable region in the form of, inter alia, another basic amine or a polar, nonbasic arrangement (e.g., amide). The basic amine portion of the pharmacophore, historically represented by a 4-substituted imidazole, is replaced by the N,N-dialkylamino motif (most commonly, a substituted piperdine or pyrrolidine or their bicyclic variations) in more recent nonimidazole analogues. Early pharmacophore modeling based on a set of imidazole antagonists suggested two hydrophobic pockets and four hydrogen-bonding sites on the receptor, two of which engage the imidazole ring.¹⁰⁴ On the basis of mutational and homology modeling studies of the H₃ receptor, interaction of the imidazole ring of H₃ agonists with aspartate 114 in transmembrane domain 3 (TM3) and glutamate 206 in TM5 was proposed.¹⁰⁵ However, this model seems to be a long way from lending practical support to the design of new ligands. Functional consequences of the single amino acid mutations vary depending on the particular ligand, assay, and assay end point (efficacy vs potency). Occasionally virtual H3 screening models have been considered;¹⁰⁶ however, the target at this point seems to be ill-suited for rational ligand design.

Despite extensive medicinal chemistry efforts waged by most major pharmaceutical companies, the overall chemical diversity of H₃ antagonists appears to be moderate, perhaps dictated by the structural requirements of the receptor. Certain structural fragments, in particular, the (*N*,*N*-dialkylamino)propoxyphenyl motif **27** (n = 1), consistently repeat themselves across different series. A bicyclic fused aryl or heteroaryl linearly linked to several monocyclic fragments is perhaps the most widespread structural arrangement among nonimidazole H₃ antagonists, although a combination of three to five linearly linked monocycles is also quite common. Very few unique structural arrangements have been reported, as will be apparent from the discussion of individual series, grouped by therapeutic indication and originator company below.

Cognitive Disorders

The use of H₃ ligands to improve cognitive dysfunction associated with a number of neuropsychiatric diseases such as AD, schizophrenia, and ADHD has been investigated both preclinically and clinically. Although these conditions most likely disrupt multiple neurotransmitter pathways, treatments have traditionally focused on a single system. As discussed above, H₃ antagonists/inverse agonists may offer a unique approach in their ability to affect multiple neurotransmitters in regions of the brain associated with memory and learn-ing.^{95,97,98,107} While multiple literature reports on the cognitive effects of histamine itself as well as other histamine receptor ligands (mediated postsynaptically via H1 receptors¹⁰⁸ and presynaptically via H_3 homo- and heteroreceptors (see Figure 2)) support the importance of the cholinergic system in signal transduction, other H₃-modulated neurotransmitters may be increasingly found to play a role. Functional consequences of the interaction between the histaminergic and cholinergic systems in the rat brain were discussed above.^{19,40-45} While elevated histamine levels in most brain regions also led to increased levels of acetylcholine, evidence of the opposite effect in basolateral amygdala in rats has been reported and attributed to activation of postsynaptic H₂ receptors, negatively coupled to cholinergic transmission.⁸⁹ Interconnection of H₃ and cholinergic signaling is also illustrated by the insensitivity of the $H_3^{-/-}$ mice to the otherwise amnesic effects of the cholinergic receptor antagonist scopolamine.⁴⁷ In addition, as suggested by literature evidence, histamine itself may also play a direct procognitive role.¹⁰⁹ Inhibition of histamine synthesis deteriorates short-term memory, while administration of exogenous histamine and histidine improves short-term memory.¹¹⁰ This is in good alignment with the observed increased output of acetylcholine from nucleus accumbens and cortex of freely moving rats, caused by exogenous histamine and blocked by H1 antagonists.^{19,40} Consistent with histamine signaling pathways, $H_1^{-/-}$ mice exhibit impaired memory characteristics, translating into reduced object recognition and maze performance.111,112 At the same time, impaired cholinergic signaling is believed to play an important role in the cognitive dysfunction aspect of schizophrenia, largely untreated by the current medications. In fact, the possibility of cholinergic signaling disruption by the very antipsychotic drug therapies currently in use has been discussed.¹¹³ Increased levels of histamine brought on by the administration of H₃ receptor antagonists may conceivably counterbalance this effect. Furthermore, both acetylcholine and histamine itself can regulate dopamine and glutamate release, abnormalities in which are also strongly associated with schizophrenia pathology.¹¹⁴ In addition, dopaminergic neurotransmission and noradrenergic neurotransmission play key roles in memory and attention and are subject to histaminergic regulation, thereby linking H₃ receptor with attentiondeficit hyperactivity disorder (ADHD).

An important role in the pathophysiology of ADHD is attributed to impaired catecholamine signaling, believed to be caused by dysfunction of the respective dopamine and norepinephrine transporter systems.¹¹⁵ Current stimulant and nonstimulant ADHD treatments (Ritalin (methylphenidate). Adderall (a mixture of dextroamphetamine and racemic amphetamine salts), and Strattera (atomoxetine)) block reuptake of dopamine and norepinephrine and/or increase their release into the neuronal synapse.¹¹⁶ In addition, however, methylphenidate and atomoxetine were also found to elevate prefrontal cortex levels of histamine in rats.¹¹⁷ Similarly, the α_1 agonist modafinil (Provigil), found to be effective in the treatment of ADHD in several clinical trials,¹¹⁸ although rejected for that particular indication by FDA in 2006, indirectly modulates the histaminergic system, in addition to its other CNS actions,¹¹⁹ and is efficacious in the treatment of narcolepsy (vide infra). It is possible then that improvements in ADHD symptoms observed with these drugs are partly due to the increased levels of histamine itself, likely acting as an integral contributor to the overall improved signaling. In that regard, an H₃ antagonist may be a valuable addition to the existing therapies.

In addition to improving cognitive aspects, the role of H_3 antagonists may extend to provide some degree of neuronal protection in such diseases as AD and PD because of, among other possible mechanisms, their inhibitory effect on the PLA₂ signaling pathway (see Supporting Information) and the accompanying generation of cell-damaging electrophilic metabolites.¹²⁰ At the same time, in an apparent contradiction to the current prevailing view, the potential role for H_3 agonists in cognitive improvement and neuronal protection was contemplated on the basis of GSK-3 β signaling pathway (see Supporting Information).

Historic H₃ antagonists, such as **5**–7 and **34** (GT-2016,¹²¹ Figure 9), have been reported to improve cognitive function, memory, spatial orientation, attention, and learning in a variety of rodent models.^{56,98–100,122–124} It is important to note, however, that performance in certain emotional memory



Figure 9. H₃ antagonists/inverse agonists for cognition.

tasks, such as those preceded by contextual fear conditioning and mediated by neural changes in basolateral amygdala, appears to be inhibited by histamine and H₃ antagonists.^{45,125} This effect aligns with the ERK signaling pathway activated by H₃ agonists¹²⁶ (see also Supporting Information) and with the pattern of histaminergic regulation of acetylcholine in the basolateral amygdala, discussed above and seemingly different from the mechanism in the cortex. Compound **5** has also recently been shown to exhibit an antipsychotic-like profile in mice¹²⁷ and improvements in learning deficits in a childhood epilepsy model.¹²⁸ Detailed summaries of in vitro and in vivo effects of brain histaminergic modulation with historic as well as more recent H₃ ligands are available in recent reviews.^{53,56,95,98,101,129}

Gliatech investigated the potential use of the imidazolebased $16^{130,131}$ for a variety of indications including ADHD, AD, insomnia, dementia, anxiety, sleep disorders, and eating disorders.⁵² The compound has a high affinity at the rat cortical H₃R ($K_i = 0.18$ nM), but a 40-fold lower affinity at the human receptor. It also binds strongly to human α_{2a} , human α_{2c} , and human H₄ receptors.^{52,99} Compound **16** was progressed into phase I clinical trials in 1998 and reached phase II enrollment in 2000 for the treatment of ADHD before the trials were halted. Subsequent to Gliatech's 2002 bankruptcy and transfer of intellectual rights to Merck, **16** was identified as a protean agonist^{32,33} in that it displays a complex pharmacological profile ranging from full agonism to full antagonism in various in vivo or in vitro systems.¹³² In contrast, recent nonimidazole H₃ ligands, displaying in vivo profiles characteristic of H₃ antagonists, have typically been reported as inverse agonists in vitro.

The FUB-UCL-INSERM-Bioproject consortium reported **35** (FUB-181) as an efficient and selective H₃ antagonist/ inverse agonist that increased turnover rates of histamine, norepinephrine, and serotonin in key brain regions^{54,133} and demonstrated beneficial learning and memory properties in rodents.⁵⁶ However, as previously mentioned, imidazole-based compounds such as **5**, **6**, and **35** are generally considered unsuitable for clinical development because of CYP450 liabilities. Stark and co-workers replaced the imidazole with piperidine, leading to tiprolisant **36** (BF-2649/FUB-649),¹³⁴ which increased dopamine and acetylcholine brain levels in rats and showed memory and learning enhancing properties in mice without observable changes in locomotor activity or hepatic toxicity. The compound also showed significant activity in several mouse models of schizophrenia.^{58,135} Compound **36** is being developed through a collaboration between Ferrer and Bioproject⁵² and is currently in phase II clinical trials for schizophrenia (begun June 2008)^{58,95,136} as well as phase II trials for narcolepsy/excessive daytime sleepiness (EDS) associated with PD (begun Oct 2007, vide infra).^{52,136} However, an independent group from Abbott has recently indicated potential development concerns including CYP2D6 inhibition, high hERG activity, and other issues.⁹⁵ Another clinical trial for a cognitive indication, a phase I ADHD study with betahistine **72** by Obecure,¹³⁶ is discussed in the section Obesity and Diabetes Mellitus.

Abbott. Abbott has been very active in the H₃ receptor field, having nominated two compounds, 37 (ABT-239, Figure 9) and ABT-834 (structure not disclosed), for clinical development and having made significant contributions to the understanding of H₃R in vitro and in vivo pharmacology.^{95,98,101} An initial screening hit, phenoxypropylpiperazine analogue 38 (A-923), was a potent binder of the rH_3R $(K_i = 2 \text{ nM})$ and served as a useful lead despite its poor bioavailability and selectivity.¹³⁷ Lead optimization led to D-alanine type derivatives **39** (A-304121) and **40** (A-317920)¹³⁸ which showed good binding in initial screening at the rH₃R $(pK_i = 8.6 \text{ and } 9.2, \text{ respectively})$ but later showed significant affinity dropoff at the hH₃R when the receptor became available ($pK_i = 6.1$ and 7.0, respectively). Both compounds were selective over H1 and H2, showed good oral bioavailability,¹³⁹ and were active in rodent cognitive models. However, 39 caused phospholipidosis, a condition clinically linked to pulmonary dysfunction, in which high partitioning of a compound into lipid bilayers causes an impairment of phospholipid turnover. This toxicity was attributed to the compound's dibasic amine and cationic amphiphilic nature¹⁰¹ and was not observed with the less basic 40. Alternatively, a biphenyl fragment was incorporated in 41 (A-320436), which showed good binding to hH_3R (pK_i = 8.8) but poor brain penetration and lack of efficacy in cognitive and ADHD models.¹⁴⁰ The biphenyl motif has also been successfully utilized in other Abbott compounds such as 73 (A-331440) and 74 (A-417022, Figure 12), which demonstrated activity in antiobesity models (vide infra). The dimethylpyrrolidine 42 (A-349821) showed good affinity and selectivity for rH_3R and hH_3R (p $K_i = 9.4$ and 8.8, respectively) and had good bioavailability and low clearance in dog and monkey. The compound was efficacious in some cognition models, but brain penetration was modest compared to other Abbott compounds (brain/plasma (B/P) of ~ 1). Since the compound was not a P-glycoprotein substrate, the reduced CNS access was attributed to its amide nature. Compound 42 was not genotoxic and showed no overt toxicity in a 2-week rat study. Unfortunately, QTc prolongation-related issues in a canine safety evaluation precluded further development. 51,55,101,141

To improve absorption and druglike properties, **37** was designed as a conformationally restricted analogue with the propyloxy spacer tied up in a benzofuran. It is potent at the rH₃R ($K_i = 0.45$ nM) and hH₃R ($K_i = 1.35$ nM) and has excellent selectivity against 80 other non-H₃ CNS receptors. Compound **37** also has good bioavailability in several species

(rat 52%, dog 74%, and monkey 89% at 1 mg/kg) and no inhibition of CYP450 enzymes. Excellent activity was observed in several rat attention, cognition, and social memory models related to ADHD and schizophrenia. Increases in acetylcholine and dopamine levels in vivo have been registered at drug doses that parallel behavioral efficacy in animal models. The 4-cyanophenyl-substituted benzofuran **37** demonstrates high brain partitioning (*B*/*P* > 34) and potency in cognition models compared to the open-chain-spacer biphenyl analogue **42**.^{95,142–144} However, high plasma protein binding and phospholipidosis have also been noted as a result of the highly lipophilic nature.^{97,101} Although a process synthesis was disclosed,¹⁴⁵ development of **37** was halted because of QTc prolongation observed in monkeys.^{52,101,146}

Additional SAR studies that explored phenyl ring substitution¹⁴⁷ and heterocyclic replacements resulted in the discovery of **43** (A-688057) and **44** (A-687136).¹⁴⁶ Pyrazolesubstituted benzofuran **43** showed good affinity for the rH₃R and hH₃R (pK_i = 8.5 and 9.3, respectively) and an improved hERG profile and was effective in preclinical cognitive models. The lower lipophilicity, compared to **37**, also suggested a lower potential for phospholipidosis. However, less promising pharmacokinetic properties (rat oral BA 26%, $t_{1/2} = 2.9$ h; dog oral BA 30%, $t_{1/2} = 1.7$ h; monkey oral BA 8%, $t_{1/2} = 1.8$ h) suggested a short half-life in humans that may be unfavorable for conditions such as ADHD.

Replacement of the benzofuran core in 37 has led to several interesting new series exemplified by 45, 46, and 47. The quinoline 45 showed good in vitro potency (rH₃R K_i = 0.93 nM, hH₃R $K_i = 0.24$ nM), PK in rat (90% oral BA, $t_{1/2} = 5.3$ h), brain penetration, and potency in cognitive models. However, the compound was abandoned because of photoinstability of its free base form and cardiovascular concerns such as increased heart contractility in dog.148 The naphthalene-based 46 also showed good potency in vitro (rH₃R $K_i = 2.5$ nM, hH₃R $K_i = 0.24$ nM) and acceptable rat PK (55% oral BA, $t_{1/2} = 5.5$ h). Notably, this analogue features an even higher B/P ratio (72) than 37.¹⁴⁹ The multikilogram synthesis of pyridazinone 47, which displays favorable biological, hERG, and PK profiles (hH₃ $K_i = 0.8$ nM, rat PK (1 mg/kg, oral (po)) of 84% BA and $t_{1/2} = 4.9$ h), has been published. ^{150,151} ABT-834 (structure not disclosed), the backup to 37, reportedly entered clinical trials for cognition in 2003, but no data have been released, suggesting that development may have been discontinued.^{52,98}

Abbott has also disclosed the natural product conessine **48** as an H₃ antagonist discovered in a high-throughput screen.¹⁵² The potent (hH₃R $K_i = 5.4$ nM) and novel steroid-based compound penetrates the CNS, but its profile suffers from a slow CNS clearance and adrenergic activity. Optimization efforts led to the *N*-methyl-D-valine analogue **49** which has favorable potency (hH₃R $K_i = 0.21$ nM), improved selectivity, and activity in a mouse behavioral model of cognition.¹⁵²

GlaxoSmithKline. Researchers at GlaxoSmithKline have developed a novel H₃ active tetrahydrobenzoazepine scaffold from initial screens of the phenoxypropylamine chemotype. The observation that the propylpiperidine motif in the initial lead **50** could be replaced with various aryl or heteroaryl ethers (e.g., **51**) led to the discovery of **52** (GSK-189254).⁹⁷ The tetrahydrobenzoazepine **52** is potent and selective at rH₃R and hH₃R (pK_i = 9.2 and 9.9, respectively), has a good pharmacokinetic profile, and penetrates the CNS, specifically binding to key brain areas including temporal cortex

preparations derived from brain tissue from severe Alzheimer's patients. The compound promotes the release of acetylcholine, norepinephrine, and dopamine in rat cortex microdialysis studies (0.3-3 mg/kg, po) and shows activity in several rodent models of cognitive performance.^{52,153} In addition, procognitive effects, receptor occupancy, and CNS exposures remained constant over multiple day twice-a-day (b.i.d.) dosing without evidence of tolerance or accumulation. Compound 52 entered phase I trials for dementia (2005, results not disclosed)^{52,58} and neuropathic pain (vide infra) and phase II for narcolepsy (vide infra).¹³⁶ In addition, ¹¹Clabeled 52 has been used in a clinical study of another H₃ antagonist, GSK-239512 (structure not disclosed), to measure brain receptor occupancy with positron emission tomography (PET).¹³⁶ Glaxo is currently recruiting for a safety and tolerability study of GSK-239512 (mild to moderate AD patients),¹³⁶ but no longer lists **52** in its pipeline (Feb 2009).⁵²

Two additional GSK compounds, the pyrazinetetrahydrobenzoazepine **53** (GSK-207040) and diazepane **54** (GSK-334429) have also been disclosed to possess intriguing activity in cognition and pain models.¹⁵⁴ Both compounds are high affinity functional inverse agonists at the hH₃R (p K_i = 9.7 and 9.5, respectively; GTP γ S pIC₅₀ = 9.2 and 8.6, respectively) and reverse memory impairment effects caused by scopolamine in rats in a passive avoidance model (0.3– 1 mg/kg, po). In addition, the potential of **53** for the treatment of schizophrenia has been evaluated. The compound significantly enhanced object recognition memory and attenuated isolation rearing-induced deficits in prepulse inhibition but did not reverse amphetamine-induced increases in locomotor activity (3 mg/kg, po), a profile described as suggesting its therapeutic utility against cognitive and sensory-gating deficits but not positive symptoms of schizophrenia.¹⁵⁵

Merck/Banyu. Merck/Banyu has disclosed a series of potent, selective, and CNS-penetrating quinazolinones including **55**, which shows reasonable pharmacokinetic profiles in multiple species without adverse cardiovascular or CNS effects in dogs (3 mg/kg, intravenous (iv)) or mice (100 mg/kg, po), and has reportedly been selected as a clinical development candidate.¹⁵⁶ The pyrrolidinylpropyloxy side chain in **55** was constrained as a cyclobutylpiperidinyloxy moiety in a series exemplified by **56**,¹⁵⁷ for which a process patent has been filed.¹⁵⁸ Banyu has also disclosed a regioisomeric quinazolinone **57** that was optimized to overcome initial hERG and adrenergic h α_{1A} activity concerns.¹⁵⁹ An H₃ antagonist, MK-0249 (structure not disclosed), is reported to be in several phase II trials for the treatment of AD, ADHD, cognitive impairment associated with schizophrenia, and excessive daytime sleepiness.^{52,95,136}

Pfizer. Pfizer has recently disclosed the structure of **59** (PF-03654746, Figure 10), an H₃ antagonist currently in phase I clinical development for the treatment of cognitive impairment in schizophrenia and AD,⁵² previously also investigated for the treatment of ADHD.¹³⁶ The compound displayed single-digit nanomolar potency at hH₃R in binding and β -lactamase reporter gene assays. Preclinically **59** enhanced histamine (drug minimal efficacious dose (MED) 1 mg/kg, sc) and acetylcholine (MED 3.2 mg/kg, sc) release in rat prefrontal cortex and demonstrated efficacy in a novel object recognition 24 h delay rat model (0.32 mg/kg, sc).^{160,161} The compound was developed from the structural lead **58** and evolved into **59** as a result of optimization of, among other parameters, physicochemical properties to avoid the phospholipidosis liability seen with early analogues. Target

engagement biomarkers in phase I clinical studies included histamine and tele-methylhistamine, measured in cerebrospinal fluid (CSF) of human volunteers after a multiple fixeddose oral study (14 days, 3 mg, once a day (q.d.)). On the basis of favorable results, **59** was advanced into and completed a phase II trial study for ADHD,¹³⁶ although the program was apparently discontinued in March 2009.⁵² Insomnia, presumably mechanism-based, was disclosed as the major observed adverse event.¹⁶⁰ Compound **59** has also completed a phase II clinical study for allergic rhinitis and is entering a phase II study for narcolepsy¹³⁶ (vide infra). Additionally, a phase I trial for AD is listed as currently active.¹³⁶

Dual-Activity Compounds from Various Companies. Several studies have identified dual-activity compounds, with which engagement of additional targets could conceivably lead to cognitive synergy with H₃ antagonism. In addition to 32^{102} discussed above, compounds with dual H₃-acetylcholinesterase inhibitory activity have been reported by Johnson & Johnson {e.g., 60 (hH₃R $K_i = 0.98$ nM; AChE $IC_{50} = 350 \text{ nM}\}^{162}$ and University of Parma {e.g., **61** (hH₃R pK_i = 8.70; AChE pIC₅₀ = 5.96)}¹⁶³ and suggested for use in AD drug therapy. The combination of HNMT inhibition and H₃ antagonist activity in **33** was discussed above.¹⁰³ Dual H₃/M₂ (presynaptic acetylcholine) receptor antagonists {e.g., 62 (gpH₃R $K_i = 18 \text{ nM}; hM_2R K_i = 0.17 \text{ nM})},$ optionally in further combination with an acetylcholinesterase inhibitor, were claimed for the treatment of AD and other cognitive disorders in a patent application, targeting an increase in acetylcholine levels via multiple mechanisms.¹⁶⁴

Sleep-Wake

As discussed above for insomnia, histaminergic modulation has a pronounced effect on sleep and wakefulness. The wakepromoting effect of histamine stems from its activation of cortical and arousal system neurons via H1 receptors.¹⁶⁵ In the sleep-wake cycle histaminergic and most other neurons release neurotransmitters and discharge at the highest rate during the waking period, decrease their activity during slowwave or nonrapid-eye-movement (NREM) sleep, and stop firing during paradoxical or rapid-eye-movement (REM) sleep, the two forms of sleep alternating with each other on a periodic basis.¹⁶⁶ Reduced CSF histamine levels have been found in human hypersomnia,¹⁶⁷ while H₃ antagonism has been shown to have wake-promoting effect through the induction of histamine release. Several studies have been undertaken to investigate the use of H₃ antagonists to treat excessive daytime sleepiness (EDS) and narcolepsy-associated uncontrollable cataplexy (sudden loss of muscle tone), believed to result from excessive somnolence and inappropriate transitions into paradoxical sleep.¹⁶⁸ Current treatments of narcolepsy include classic CNS psychostimulants such as amphetamine and caffeine, whose efficacy is hampered by issues of tolerance and increased locomotor activity, and the α_1 agonist modafinil, which affects wakefulness through an indirect activation of the histaminergic system. 62,63,99,169 Classic H_3R antagonists such as 5, 7, 16, and 36 have been shown to increase wakefulness and decrease sleep in rodent, feline, and canine models. Notably, the effects are absent in knockout mice lacking either the H_3 or histidine decarboxylase gene. 58,62,63,97,98,135,170 Compound **36** promoted wakefulness, decreased abnormal direct-rapid-eye-movement (DREM) sleep, and reduced EDS in orexin knockout $(Ox^{-/-})$ mouse



Figure 10. H₃ antagonists/inverse agonists for cognition.

model of narcolepsy as well as in two small human clinical studies.^{171,172} Orexins are neuronally released excitatory peptides promoting wakefulness and closely linked to aminergic neurotransmission. Differential effects of acute and repeat dosing of the H₃ antagonist 52 on sleep-wake cycle and narcolepsy have been described. Significant reduction of the wakefulness-promoting effect and increased suppression of narcoleptic episodes in $Ox^{-/-}$ mice have been reported with subchronic dosing (8 days, 10 mg/kg, b.i.d., po).¹⁷³ Tetrahydrobenzoazepine 52 also increased wakefulness in rats and was advanced to phase II clinical study for narcolepsy, although the trial was terminated.^{58,136} As discussed above, insomnia was reported as the major adverse event in a phase II clinical study for ADHD with **59**, which is currently entering a phase II study for narcolepsy.¹³⁶ The issue of higher drug dose, generally needed for robust wake effect compared to cognitive effects, was addressed in a recent wake-promoting study of 5, 7, 37, and 52, which demonstrated that relatively low levels of wake activity linearly correlated with receptor occupancy up to 80%. In contrast, a dramatic nonlinear increase in waking activity at higher receptor occupancy levels (>80%) was observed, potentially suggesting an altered mechanism due to a substantially modified overall neurotransmitter profile.174

Johnson & Johnson. In addition to the seminal efforts in the H_3R field, including the cloning and functional

expression of the human receptor in 1999.⁶ researchers from Johnson & Johnson have contributed extensively to the development of H₃R antagonists with wake-promoting and procognitive characteristics. Identified in a highthroughput screen, the calcium channel blocker 63 (RWJ-20085, also JNJ-280566, Figure 11) showed micromolar hH₃R activity.¹⁷⁵ Reminiscent of the natural product aplysamine, 63 exemplified the (N,N-dialkylamino)propoxyphenyl motif 27. Early optimization efforts of this imidazopyridine provided 64 (JNJ-6379490), which displayed potent hH₃R binding ($K_i = 2 \text{ nM}$) and excellent selectivity over other histamine receptors. Compound 64 displayed good oral bioavailability in the rat and dog as well as high brain penetration. Increased wake time was observed in rats (0.6 mg/kg, sc), although the activity appears to be limited to the first 2 h, even at high doses (10 mg/kg). Imidazopyridine 64 also demonstrated activity in a rat short-term memory model, although development was apparently halted because of idiosyncratic toxicity in the dog.^{170,175}

Further work led to the 4-(aminoalkoxy)benzylamine series typified by the neutral antagonist **65** (JNJ-5207852), which showed good potency (hH₃R, $pK_i = 9.24$), selectivity, CNS penetration, and oral biovailability. This candidate also promoted wakefulness in rodents (10 mg/kg in the rat) and decreased NREM/REM sleep without rebound hypersomnolence.





No effects on sleep or wakefulness were observed in $H_3^{-/-}$ mice. In addition, **65** decreased episodes of cataplexy in a genetically narcoleptic Doberman Pinscher model and showed efficacy in learning and memory tests without affecting locomotor activity. Unfortunately, **65** shared the liabilites of related diamines, including an unfavorably high volume of distribution (> 100 L/kg), long plasma half-life (15–17 h), and brain residency in the rat (full receptor occupancy in the striatum observed 24 h after 30 mg/kg po dose), as well as phospholipidosis-related toxicity, precluding further development.^{55,170,176,177} Conformational restriction of the aminoalkoxy chain provided compounds such as **66** (JNJ-7737782) which was active in rodent models of wakefulness (1 mg/kg, sc).¹⁷⁸

Further examples of Johnson & Johnson's diamines with rigidified linkers include alkynes exemplified by **67** (JNJ-10181457) with good H₃ potency, selectivity, brain residency, and pharmacokinetic profiles.^{52,170} Phenylacetylene **67** promoted wakefulness in mice and reduced cataplectic attacks in narcoleptic dogs.¹⁷⁹ The compound also increased noradrenaline and acetylcholine levels in rat frontal cortex without affecting dopamine and serotonin and showed activity in learning and memory models without locomotor effects (10 mg/kg, intraperitoneal (ip)).^{128,170,180} Given the wakepromoting action of **67**, its differential effect on neurotransmitter levels is noteworthy and may indicate their respective critical or flexible roles in the maintenance of arousal state.

JNJ-17216498 (structure not disclosed) is an orally active and selective H₃ antagonist that has completed a phase II trial for narcolepsy.¹³⁶ The compound was reportedly active in preclinical narcoleptic models and showed a favorable pharmacokinetic profile in phase I.^{52,97} The syntheses and crystal structures of the centrally substituted imidazole **68**¹⁸¹ and pyrrolidinopropoxyphenylquinazoline **69**¹⁸² have also been recently highlighted, suggesting continuing interest in these series.

Researchers from Johnson & Johnson have also investigated a series of dual H₃ antagonist-serotonin reuptake

inhibitors to address fatigue, somnolence, and cognitive impairment frequently diagnosed in patients suffering from depression. Wake-promoting agents such as modafinil have been successfully used with selective serotonin reuptake inhibitors (SSRI) to treat excessive sleepiness.¹⁸³ The notable feature of serotonin in the brain is its dual role in the sleepwake cycle. It appears that initial rapid arousal action is followed by a delayed increase in NREM sleep as a result of the stimulated synthesis and release of sleep-promoting factors.¹⁸⁴ This effect appears to manifest itself in the daytime sedation observed with SSRI (e.g., fluoxetine) and may be targeted for neutralization by H₃ antagonists. Fusion of H₃ and SSRI pharmacophores has been achieved in the structure of **70** (JNJ-28583867, $hH_3R K_i = 11 nM$, hSERT $K_i = 4 \text{ nM}$), which increased serotonin, norepinephrine, and dopamine levels in rat brain (hNET $K_i = 121$ nM, hDAT $K_i = 102$ nM). The compound shows antidepressant activity in mice and promotes wakefulness in rats.^{185,186} As could be expected from its effect on serotonin, 70 decreased NREM and suppressed REM sleep ($\geq 1 \text{ mg/kg, sc}$).

Pain/Analgesia

The potential role of H_3R ligands in the treatment of pain remains unclear based on the conflicting data for H₃R agonists and antagonists in pain models. As discussed above for H₃ agonists, the effects of histamine itself as well as other H₃ ligands on nociception are highly dependent on the conditions of the specific pain model. In the absence of clear understanding of mechanisms involved, the practical issue of relative receptor selectivity adds an extra variable to the already complex picture, as mediation of nociception by the H₄ receptors has also been reported.¹⁸⁷ For example, while the moderately H₃ vs H₄ selective antagonist 5 (rH₃R p $K_i = 8.4$, $hH_3R pK_i = 7.3, rH_4R pK_i = 7.3, hH_4R pK_i = 6.9$ ^{8,188} demonstrated moderate activity against acute nociceptive pain in some studies,⁸⁷ in other cases it had no effect by itself and also reduced the antinociceptive effects of morphine (tail-immersion and hot-plate tests and acetic acid-induced

writhing).¹⁸⁹ In contrast, the H₃ agonist 1 potentiated the effects of morphine. Antinociceptive activity for the H₃R agonists 1, 4, and 13 has been reported in some models, while in other studies 3 caused allodynia and 1 was hyperalgesic, preventing the effects of 5.58,79,85,99,190 Importantly, the limitations of these compounds as in vivo H₃ tools, including poor CNS penetration and rapid metabolism of 1⁸⁵ and H₄ agonist activity of **3** and **4** (hH_4R pEC₅₀ = 7.9 and 7.8, respectively),¹⁸⁸ complicate the interpretation of data. There is also evidence that the effect of an H₃ antagonist in a given model may even be dependent on the delivery route. Thus, central (intracerebroventricular (icv)) administration of 5 increased while systemic (ip) administration decreased pain threshold in the rat sciatic nerve ligation model of neuropathic pain.¹⁹¹ In the same study the very nature of histamine's effect was dose-dependent, with low icv dose decreasing and high icv dose increasing the nociceptive threshold, highlighting again the potential challenges of H₃-based approach to the treatment of pain.

Overall, however, it appears that H₃ antagonists are consistently more useful under the conditions of neuropathic pain, in which peripheral chronic inflammation or nerve injury and continuous CNS stimulation cause plastic changes in pain descending pathways with the dominance of facilitatory mechanism over inhibitory.⁸⁹ As a pathology of disregulated neurotransmission, neuropathic pain appears a possible if yet unproven target of H₃ modulation. This hypothesis has been given further support by the discovery of several series of selective H₃ antagonists at Glaxo, including the previously discussed tetrahydrobenzoazepines 52 and 53 and diazepane 54, which have demonstrated a reduction of mechanical hyperalgesia and allodynia in a chronic constriction injury and varicella zoster virus rat models of neuropathic pain.^{154,192} In addition, specific H₃ receptor binding sites were established in the human dorsal horn of the spinal cord and dorsal root ganglion, the key relay center for ascending and descending pain conductance. Although unknown, the mechanism of action is hypothesized to be similar to that of the neurotransmitter reuptake blockers duloxetine (Cymbalta) and venlafaxine (Effexor), which increase brain levels of monoamines such as serotonin and norepinephrine.^{192,193} Compound 52 was in phase I clinical investigation in an electrical hyperalgesia translational model of neuropathic pain.¹³⁶

Epilepsy

The involvement of histamine in seizures has long been recognized based on the proconvulsant properties of traditional H₁ antihistamines.¹⁹⁴ Histamine and agents that raise histamine levels, such as histidine and the HNMT inhibitor metoprine, have shown potent anticonvulsant effects most likely through H₁-mediated excitation of interneurons and inhibition of hippocampal principal neurons. Consistent with this observation, the H₃ antagonists 5 and 6 have shown protective anticonvulsant effects in various animal models based on their facilitation of brain histamine release. H₃ agonists have also been shown to prevent the protective effect of H₃ blockers.^{99,100,195} No clinical data are available with the exception of a small uncontrolled trial with 36 (12 patients), in which anticonvulsant effects were observed.¹⁷² At high doses, the H₃ antagonists 5, 7, 16, 40, and 42 have shown proconvulsant activity in animal models; however, the mechanism of this biphasic effect is unknown.^{138,141}

Obesity and Diabetes Mellitus

Obesity is mainly characterized by the accumulation of an excessive amount of body fat. Globally, more than one billion adults are overweight with a body mass index (BMI) over 25 and more than 300 million are clinically obese with a BMI over 30. The medical consequences of this condition are serious and associated with a significant decrease in life expectancy. It is well established that even modest weight loss reduces the medical risk for the development of type 2 diabetes, cardio-vascular disease, and cancer in the obese population. Although obesity can and has been targeted via multiple biochemical pathways, the resilience of the compensatory mechanisms against weight loss (due to multiple redundant regulatory circuits aimed at maintaining the status quo) has been a formidable problem.

The key to combating obesity and ensuing downstream pathologies appears to be in the restoration of the energy intake/expenditure balance. Appetite and food intake are centrally regulated by the hypothalamus through a number of pathways involving or xigenic and anorexigenic signaling peptides and neurotransmitters acting at the appropriate receptors.¹⁹⁶ The combined regulatory signals at the state of satiety were designed by nature to promote a decrease in food intake and increase in energy expenditure, supplied by increased glucose metabolism and lypolisis in adipose tissue. The efficiency of signaling, however, may be severely disrupted in states of pathology, including obesity, requiring exploitation of additional mechanisms different from those already compromised. Among the most visibly disregulated in the condition of obesity and often accompanying type 2 diabetes are the signaling pathways of insulin and leptin, the latter being the most prominent among anorexigenic signaling peptides, released by adipocytes and circulating at higher levels in the satiety state.

In addition to the regulatory action from neuropeptides, appetite and food intake effects also appear to be subject to regulation by aminergic neurotransmitters, including dopamine, norepinephrine, serotonin, and histamine. Significant work has been done demonstrating that histamine stimulates postsynaptic H₁ receptors to inhibit feeding. In 1973 it was first shown that histamine suppresses food intake when intraventricularly injected into cats.¹⁹⁷ Likewise, acute or continuous administration of histamine into the lateral ventricle or the suprachiasmatic nucleus of the hypothalamus in rats reduced food intake.^{198,199} Intraperitoneal injection of histamine enhancing agent L-histidine, or the HNMT inhibitor metoprine, has been shown to have the same food-reducing effect,^{200,201} while intracerebroventricular administration of α -fluoromethylhistidine 22 (Figure 7), an HDC inhibitor, significantly increased food intake in rats by depleting histamine.²⁰² Administration of the H₁ receptor agonist 2-(3trifluoromethylphenyl)histamine 71 (Figure 7) depressed food intake in wild-type (WT) rats,¹⁹⁹ whereas many studies have demonstrated that H1 antagonists induce feeding.²⁰³ In particular, treatments with tricyclic antidepressants and atypical antipsychotics often induce body weight gain which has been proposed to be potentially mediated by H₁ antagonism.²⁰⁴ The degree of increase in food intake in rodents treated with tricyclic antidepressants was related to these compounds' H_1 receptor affinities.²⁰⁵ This fact is also in agreement with data in humans, where use of atypical antipsychotic drugs, such as clozapine and olanzapine (Zyprexa), results in untoward orexigenic effects arising from central H1 antagonism

 $(K_d(hH_1R) = 3.1 \text{ nM} \text{ and } 0.087 \text{ nM}, \text{ respectively}).^{206}$ Similarly, unwanted body weight gain in humans was observed as part of treatment of allergic conditions with H₁ antihistamines. However, this effect is significantly diminished with the second generation antihistamines because of lower brain penetration.²⁰⁷

Consistent with these findings are the data from the rodent knockout models. $H_1^{-/-}$ mice on a high-fat diet have been reported to accumulate significantly more fat than WT animals, although no statistical difference in total body weight or food intake between the two groups was observed.²⁰⁸ In another study histamine-deficient HDC^{-/-} mice on a high-fat diet developed significant difference in body weight relative to standard-diet group after only 2 weeks, compared with 5 weeks for WT mice. Furthermore, after 8 weeks WT mice on the high-fat diet reduced their caloric intake, whereas HDC^{-/-} mice did not.²⁰⁹ Paradoxically, $H_3^{-/-}$ mice demonstrated a mildly obese phenotype characterized by increased adiposity and food intake, reduced energy expenditure, and increased resistance to insulin and leptin.⁴⁶ While the orexigenic effects were not easily explainable and were tentatively attributed to the impairment of histamine synthesis as a result of unchecked histamine release, only a 15% reduction in the histamine hypothalamus levels in $H_3^{-/-}$ mice (along with 135% increase in tele-methylhistamine levels) was observed. Alternatively, reduced H₁ receptor expression was invoked, and it is conceivable that other mechanisms of H1 receptor desensitization by higher levels of histamine, unmitigated by negative regulation via H₃, may also play a role.^{210,211}

Despite the established anorexigenic role of histamine, the nature of coupling between H₃ receptors, histaminergic tone, and the overall effect on food intake and body weight has been a recurring topic of debate. The therapeutic significance of H₃ receptor stimulation vs inhibition for feeding and weight gain appears to be a subject of more controversy than in the case of cognitive indications. Significantly different results have been reported depending on experimental paradigm, administration route, and species/strain used. For example, the H₃ antagonist 5 reduced food consumption and/or appetite in normal and in Zucker obese rats that lack functional leptin receptors.^{212,213} While a majority of other reports also link H_3 antagonism/inverse agonism to anorexigenic effects based on the expected increase in histamine tone (vide infra), these data are not entirely uniform. It was suggested in a 2006 study supported by $H_3^{-/-}$ mouse data that H_3 receptors may regulate food intake in rodents independently from modulation of histaminergic tone or subsequent changes in the activity of H1 receptors.²¹⁴ Chronic dosing with the H3 agonist 3 reduced food intake, body weight, fat mass, hyperleptinemia, and hyperinsulinemia in diet-induced obese (DIO) mice, while the opposite effects were observed with the H₃ antagonist 5. The food- and weight-reducing effects of 3 were negligible in DIO $H_3^{-/-}$ mice and augmented upon intracerebroventricular administration in WT mice, strongly implicating central H₃ receptors in the observed anorexigenic effect. While absolute brain histamine levels increased following treatment with 3, this was attributed to its accumulation in nerve terminals rather than neuronal synapse and was apparently supported by the fact of reduced tele-methylhistamine levels. Presumably independent of histamine tone, the effect of 3 was tentatively attributed to its postsynaptic H₃ effect (occasionally invoked in this case and others but hardly evidenced in a credible way)³⁸ or a presynaptic H_3 effect on release of other appetite-affecting neurotransmitters, such as dopamine and norepinephrine. These findings may offer some rationale for the obese phenotype of the $H_3^{-/-}$ mice discussed above.⁴⁶ The antiobese effects of H_3 agonists were also confirmed with 1, while intracerebroventricular administration of 5 in rats produced orexigenic effects similar to those observed in mice.²¹⁴

At the same time other studies by Banyu suggested strong species dependence of H_3 -mediated food intake and body weight effects. Thus, the same H_3 inverse agonists (not identified) produced orexigenic effects in C57BL/6J DIO mice and anorexigenic effects in Zucker obese rats, underscoring potential differences in physiology and pharmacology in rodents and emphasizing the importance of metabolic syndrome data in nonhuman primates (vide infra).²¹⁵

Contributions of H₃ receptors to the expenditure component of the intake/expenditure balance of energy homeostasis are less clear. For the safe increase in total energy expenditure the most promising target of pharmacological intervention in rodents and humans is believed to be adaptive thermogenesis, in mammals largely regulated in brown adipose tissue (BAT). Increase in thermogenesis has been linked to the up-regulation of the mitochondrial respiratory chain components as well as uncoupling proteins (proton transporters shifting the balance from ATP synthesis in mitochondria to heat generation) under control of a number of transcription regulators. Among these is peroxisome proliferator-activated receptor- γ coactivator 1 (PGC1), induced by cold exposure and β_3 adrenoceptor agonists in BAT and skeletal muscle.²¹⁶ Consequently, the effect of β_3 action of catecholamine neurotransmitters (linked to H₃ and histamine and released by the sympathetic nervous system) is increase in thermogenesis.²¹⁷ It is interesting to note that β_3 agonists affect PGC1 levels via cAMP-dependent activation of protein kinase A, a major signaling pathway of H₃ antagonists (see Supporting Information).

Obesity increases the risk of developing other characteristic conditions of metabolic sydrome such as type 2 diabetes, a disorder characterized by abnormally high blood glucose levels not caused by lack of insulin. It is believed to be the result of disregulated hormonal CNS signaling, with insulin and leptin being the primary compromised mediators, coupled with the loss of peripheral sensitivity to insulin.²¹⁸ Leptin and insulin signaling in hypothalamus is supplemented by the action of neurotransmitters, including H₃-mediated dopamine and serotonin, and directly influences glucose homeostasis. It is clear that reduction of body weight in rodents by histamine has favorable effects on such parameters as serum glucose and insulin;²¹⁹ however, it is less apparent whether these effects can be achieved independently of weight loss. Experiments in nonobese animal models of type 2 diabetes^{220,221} may answer this question and delineate the prospects for H₃ antagonists in the treatment of type 2 diabetes not associated with excessive body weight and often found in Asian population. While molecular pathways of glucose homeostasis regulation by histaminergic system are not clear, there is substantial evidence that it acts in part as a downstream effector of leptin regulation of energy homeostasis and glucose metabolism.²²² However, one challenge in emphasizing a particular mechanism, such as H₃ antagonism, lies in the potentially opposite effects of the same signaling pathways in the CNS and in the periphery. While peripheral activation of cAMP-protein kinase A pathway (linked to H₃ antagonism and discussed in the Supporting Information attachment) is



Figure 12. H₃ antagonists/inverse agonists for obesity and diabetes.

believed to increase insulin sensitivity in the skeletal muscle, its activation in the CNS increases food intake.²¹⁸ Both insulin and leptin were shown to inhibit the cAMP–protein kinase A pathway in the hypothalamus, which would seem to align them with H₃ agonist rather than H₃ antagonist or histamine-via-H₁ signaling patterns.^{218,223} Clearly, additional information may be helpful in supporting the case for H₃ antagonists in diabetes. In addition, other components of metabolic syndrome, such as liver disease and hyperlipidemia (vide infra), have been considered in conjunction with H₃ antagonism.

Challenges to realizing an H₃-based approach to the treatment of obesity and diabetes are abundant. In addition to inherent species differences discussed above, the apparent dichotomy of H₃ compounds with metabolic syndrome as opposed to cognition enhancing activity has been highlighted.¹⁰¹ Technical issues, likely common to all obesity programs, include difficulties in designing an animal model in which a decrease in feeding and/or body weight is exclusively due to the targeted pharmacological action of the drug rather than toxicological or any other side effects interfering with animal eating routine. In general, reliable extrapolation of such fundamental activity as feeding from animals to the evolutionally more accomplished humans remains a significant challenge. Despite these obstacles, several companies appear to be pursuing metabolic syndrome indications with H₃ antagonists. A satiety effect was observed with compound 36, under development by Ferrer and BioProject for neurological disorders, when administered to six human volunteers also treated with olanzapine.¹⁷² Progress of other development programs is described below.

Obecure. Obecure is developing betahistine 72 (OBE-101 or histalean, Figure 12), an H₁ agonist/H₃ antagonist for the treatment of obesity and antipsychotic medication-induced weight gain, and has considered it as a potential adjunct to statin therapy for improving blood lipid profiles.¹³⁶ Compound 72 was previously approved for human use for the treatment of Meniere's disease (vide infra) and is an orally active brain-penetrating analogue of histamine. Both molecules contain an ethylene linker between a nitrogen heteroaromatic ring and an sp³ amine. However, 72 has a complex pharmacological profile; it exhibits a combination of weak affinity (double-digit micromolar rodent K_i) and partial agonism (relative to histamine) at the H₁ receptor with somewhat higher affinity (single-digit micromolar rat K_i) and antagonism at the H₃ receptor.^{224,225} In vivo **72** reduced food intake in pygmy goats and rats.^{226,227} Several standalone phase II obesity trials and several phase II trials to assess ability of 72 to prevent weight gain in patients taking

olanzapine were commenced in 2007–2008.¹³⁶ Despite moderate receptor affinity, proposed human doses in efficacy trials do not seem to exceed 100 mg/day. However, in a 12-week clinical trial **72** failed to induce significant weight loss in obese patients (except in women younger than 50 years of age) at doses up to 24 mg b.i.d.²²⁸ Separately, Obecure is recruiting for a phase I ADHD study with **72** with planned doses of up to 200 mg/day.¹³⁶

Schering-Plough. SCH 497079 (structure not disclosed) is an H₃ receptor antagonist investigated by Schering-Plough for the treatment of obesity and diabetes mellitus. A phase II study to evaluate effect of SCH 497079 on weight in obese and overweight subjects and a phase I study to evaluate effect of SCH 497079 on metabolic parameters in subjects with type 2 diabetes mellitus began in 2008.¹³⁶

Abbott. While neither 42 nor the conformationally restricted analogue 37, discussed in the section Cognitive Disorders, appeared to have activity in animal models of obesity, continuation of the biphenyl motif with an added basic center in the form of a dimethylamino group at the 3-position of the pyrrolidine ring resulted in compounds that promoted food intake reduction and body weight loss. Abbott has investigated compound 73 (A-331440), which produced effects on metabolic parameters in DIO mice.²²⁹ In a 28 day study the high dose of 73 (15 mg/kg, po, b.i.d.) reduced weight and leptin and normalized insulin tolerance to the levels observed in lean mice. Importantly, the weight loss was largely due to specific loss of abdominal and subcutaneous fat and was not generalized across tissues. Because of the genotoxic liability of 73, Abbott developed nonclastogenic fluorinated analogues 74 (A-417022) and 75 (A-423579) with in vitro H_3 profiles similar to that of 73 and demonstrated their in vivo activity in obese rodents.^{212,230} However, Abbott has not reported a clinical development program for a metabolic syndrome indication.

Novo Nordisk. TransTech Pharma, under license from Novo Nordisk, is investigating H₃ histamine antagonists for the treatment of metabolic disorders, presumed to include obesity. By screening arrays of monoacyldiamines, Novo Nordisk discovered 1-alkyl-4-acylpiperazines as a new class of nonimidazole H₃ antagonists.²³¹ Within this series compound **76** (NNC 38-1049, hH₃R $K_i = 1.2$ nM) produced significant reduction in food intake and body weight in a 2-week study in DIO rats.²³²

Exploration of other diamines led Novo Nordisk to the cinnamic amides of (S)-2-(aminomethyl)pyrrolidines.²³³ In this novel class of compounds the two nitrogen atoms of the diamine part of the molecule are separated by a two-carbon

linker, similar to the acylpiperazine series. Selection of *S*-stereochemistry and introduction of the 4-trifluoromethyl substituent on the phenyl ring led to the discovery of 77 (NNC 38-1202), a potent and selective H₃ antagonist (hH₃R $K_i = 6.7$ nM). In line with the increased paraventricular histamine levels observed in normal rats (339% after single 15 mg/kg oral dose), 77 significantly reduced food intake, body weight, and plasma triglyceride levels in DIO rats.²³⁴ In addition, 77 showed good affinity for pig and rhesus monkey H₃ receptors (pH₃R $K_i = 18$ nM; mH₃R $K_i = 6.3$ nM) and strongly reduced average caloric intake in both species (perhaps, unnervingly so, by 75% at 1 mg/kg dose in the monkeys, although no adverse effects were observed), demonstrating for the first time the anorectic effect of an H₃ antagonist in higher mammals.²³⁵

Allergic Rhinitis

Prior to the discovery of the H₃ receptor and consideration of H₃ agonists for minimization of histamine inflammatory effects through reduction of its extracellular levels (discussed above), postsynaptic histamine H₁ receptor antagonists had already been widely used for the symptomatic treatment of allergic inflammatory conditions, and it is the apparent promise of H₄ receptor antagonists to do so in the future.⁵ Among different allergic conditions, the most pronounced response to treatment with antihistamines has been observed in the case of allergic rhinitis, characterized by nasal congestion, rhinorrea, sneezing, and pruritus and occurring in sensitized individuals upon exposure to allergens such as ragweed, grass, tree pollen, and animal dander. The allergic cascade in general is initiated by the uptake of allergens and their presentation to naive T cells, leading to their differentiation toward the T_H 2-cell phenotype.²³⁶ T_H 2 cells up-regulate expression of a number of cytokines, including IL-4, IL-9, and IL-13, leading to IgE synthesis (constituting allergic sensitization) and, upon subsequent allergen exposure, to recruitment of mast cells and basophils, binding and cross-linking of IgE, degranulation of cells, and release of inflammatory mediators, including leukotrienes, prostaglandins, cytokines, and histamine. Further downstream effects arise from actions of these mediators on tissue cells, such as epithelial, endothelial, and smooth muscle cells, leading in the case of histamine to mucus production, increased vascular permeability, and bronchospasm. Additionally, mediators' signaling back to the immune cells (eosinophils, B and T cells) further prompts their stimulation, recruitment, and polarization. It is believed that release of initial mast cell mediators, including histamine, is responsible for the generation of acute symptoms, whereas further induction of cytokines and immune cell infiltration sustain the inflammatory environment. As histamine is just a small part of the overall inflammatory cascade, antihistamines are efficacious in some but not all conditions with allergic etiology. For example, they appear to lack efficacy against asthma, despite the similarity between bronchoconstrictive effects of histamine and asthma pathology. In contrast, while histamine also plays a crucial role in the development of allergic rhinitis symptoms,²³⁷ those, with the exception of nasal congestion, appear to be well controlled by H_1 antagonists.

Lack of efficacy against nasal congestion appears to arise from a separate, H_3 -mediated, inhibitory effect of histamine on norepinephrine release and the resulting disruption of normal vascular tone. Consequently, the nonsedating antihistamines, such as loratadine and cetirizine, are often combined with α -adrenergic agonists to achieve full decongestive effect. While use of α -adrenergic agonists is limited by their systemic cardiovascular and CNS stimulatory effects, a peripherally acting H_1/H_3 combination would appear to offer a viable alternative. The vasoconstrictor effect of H₃ antagonists due to the reestablished release of norepinephrine could restore the normal vascular tone in the nose. The H₃ receptor was shown to modulate sympathetic control of nasal blood flow and nasal resistance in the cat.²³⁸ However, the action of H₃ antagonists alone is not expected to deliver a full decongestive effect in allergic rhinitis because of the need to block H₁-mediated mucus production and plasma extravasation contributing to nasal congestion. Administration of the H1 antagonist chlorpheniramine or desloratadine in combination with the H₃ antagonist 5 or 6 was shown to block the decrease in cat nasal cavity volume caused by mast cell degranulator compound 48/80. Neither an H_3 nor H_1 antagonist was efficacious when administered alone.²³⁹ A drug with dual H_1/H_3 antagonist properties or an H₁/H₃ antagonist combination with minimal brain penetration could prove useful for the treatment of nasal congestion and avoid the hypertensive liability of current therapies.

GlaxoSmithKline. Glaxo is developing two dual H₁/H₃ antagonists, GSK-835726 and GSK-1004723, for the treatment of allergic rhinitis. By early 2009 GSK-835726 had been advanced to a phase II environmental challenge chamber efficacy study, which it completed several months later.¹³⁶ The compound was evaluated against the H₁ antagonist cetirizine and placebo in a crossover trial at single oral doses up to 100 mg. In the absence of direct structural information, we believe that GSK-835726 may have structure **78** (Figure 13). In 2006 Glaxo disclosed **78** in a single-compound patent.²⁴⁰ The human H₃ and H₁ receptor affinities (p*K*_i) for **78** are 9.5 and 5.6, respectively, and oral bioavailability exceeds 50% with a plasma half-life ($t_{1/2}$) of 2 h in rats and 90% with a plasma half-life of 5 h in dogs. In addition, **78** showed low CNS penetration in rats.

GSK-1004723 reached and completed a similar phase II chamber efficacy study in 2009, where it was compared against placebo at single intranasal doses up to 1000 mg.¹³⁶ Again, the structure has not been directly disclosed, but we believe GSK-1004723 may have structure 79. In 2007 Glaxo filed a patent application covering a novel class of 4-benzylphthalazinone dual H1/H3 antagonists.241 Compound 79 was reported to have H_1 receptor affinity (p K_i) of 7.8 and H_3 receptor affinity of 9.6. Compound 79 fully reversed histamineinduced nasal congestion in conscious guinea pigs for a period of 24 h following intranasal administration of 0.05 mg dose (a more robust effect compared to the H₁ antagonist azelastine) and exhibited lower CNS penetration in rats. Phtalazinone 79 is a clearly discernible combination of the common ((N,N-dialkylamino) propoxyphenyl) H₃ pharmacophore and H₁ pharmacophore mirroring the structure of azelastine 80 (Figure 7).

Glaxo is also investigating another dual H₁/H₃ antagonist, preclinical candidate **81** (GSK-541636, hH₃R p $K_i = 7.3$, hH₁R p $K_i = 7.9$).²⁴² Compound **81** demonstrated in vivo antiinflammatory activity in the wheal and flare guinea pig model with ID₅₀ of 0.6 mg/kg (iv) and 2.8 mg/kg (po). Oral bioavailability of 60% with a plasma half-life of 3 h in dogs and low CNS penetration in rats were disclosed. Glaxo observed that the methyl-disubstituted piperidine was the optimal amine component of the (*N*,*N*-dialkylamino)propoxyphenyl motif in this structural series. Brain penetration was reduced with the introduction of the carboxylic acid functionality,



Figure 13. H₃ antagonists/inverse agonists for allergic rhinitis.

while ethylene linker improved bioavailability relative to a previous compound that contained oxymethylene linker.

Pfizer. A phase II clinical study to test the decongestive activity in allergic rhinitis of the previously discussed phenyl-cyclobutanecarboxamide **59** was completed in August 2008 (**59** as a stand-alone treatment against Allegra (fexofenadine), Allegra-D (fexofenadine–pseudoephedrine), and placebo).¹³⁶ No results of the study have been published, and the indication has been absent from the company's pipeline since September 2008, possibly suggesting a negative outcome.⁵²

Schering-Plough. Researchers at Schering-Plough were the first to demonstrate decongestive activity of H_3 antagonists in an animal model of nasal congestion when administered in combination with an H_1 antagonist.²³⁹

Imidazole-based analogue **82** (SCH 79687) was reported as a potent, selective, and orally active H₃ receptor antagonist with decongestive activity and absence of blood pressure effect in the cat when coadministered with the H₁ antagonist loratadine.²⁴³ In addition, dual H₁/H₃ receptor antagonists, exemplified by **83** (gpH₃R $K_i = 15$ nM, rH₁R $K_i = 7$ nM), a histamine homologue–H₁ antagonist chlorpheniramine **84** (Figure 7) structural hybrid, have been reported.²⁴⁴

Ménière's Disease

Ménière's disease is an abnormality of the inner ear characterized by spontaneous attacks of vertigo or severe dizziness, fluctuating hearing loss, tinnitus (ear noises), and the sensation of pressure or pain in the affected ear. This disorder usually affects only one ear and is a common cause of hearing loss. The symptoms of Ménière's disease are associated with a change in fluid (endolymph) volume within the membranous labyrinth which is necessary for hearing and balance. An increase in endolymph can cause the membranous labyrinth to balloon or dilate, a condition known as endolymphatic hydrops. On the basis of a recent study, NIDCD (National Institute on Deafness and Other Communication Disorders) estimates that there are currently 615 000 individuals with diagnosed Ménière's disease in the United States and 45 500 newly diagnosed cases each year. The causes of Ménière's disease are not known. Some researchers believe that the symptoms are the result of a blood vessel pressing upon a nerve. Other researchers view this as a separate disease. Other investigators believe that the symptoms are the result of an autoimmune condition or an infection generated by a virus such as the herpes simple virus (HSV).

Dual H_1 agonist/ H_3 antagonist **72** (Figure 12), discussed above in the section Obesity and Diabetes Mellitus, was first registered in Canada in 1968,²⁴⁵ is currently approved worldwide for the treatment of Ménière's disease, and is marketed by Solvay Pharmaceuticals (now part of Abbott) in more than 90 countries (U.S. excluded) under the brand names Serc, Betaserc, and several others. While no branded form has been approved for sale in the U.S., **72** is available from compounding pharmacists. In the U.K., **72** is prescribed by 94% of otolaryngologists for this indication, and it is estimated that over 130 million people have been treated worldwide. Analysis on the efficacy of **72** suggested reduction of vertigo and some reduction of tinnitus in most trials. However, no effect on hearing loss was observed.²⁴⁶

The mechanism of action of **72** in Ménière's disease is not clear. A beneficial effect of histamine upon intravenous administration in patients with Ménière's has been described.²⁴⁷ A possible explanation could be the previously mentioned histamine-mediated increase in vascular permeability possibly leading in this case to drainage and a decrease of the endolymph fluid volume in the inner ear labyrinth. This effect could be produced by **72** via up-regulation of histamine levels. In another argument, the vasodilatory effect of histamine resulting in the improved blood flow to the inner ear has been invoked. This hypothesis is supported by the fact that different vasodilators have a beneficial effect in the treatment of Ménière's and by the earlier findings of increased blood flow in monkeys and dogs upon administration of **72**.^{248,249}

Cancer

Histamine and its receptors play important roles in cell proliferation in brain, colon, and breast cancers.^{250–252} Elevated expression of histamine receptors in cancer cells has been observed, suggesting that they could potentially be exploited with the goal of decreasing cellular growth and proliferation. High histamine biosynthesis and content have been reported in different human neoplasias including breast cancer, as well as in experimental tumors induced in rodents.²⁵³

In breast carcinomas, a higher expression of the H₃ receptor was observed in tumor cells relative to nontumoral tissue in the same patient. Histamine and other H_3 agonists (1 and 3) were shown to increase proliferation of human breast cancer cells at low concentrations (0.001–0.01 μ M), an effect that was blocked by the H₃ antagonists 5 and 65^{254} At the same time, no effect was observed with H₃ antagonists alone $(10 \,\mu\text{M})$. At higher concentrations, histamine strongly inhibited cell proliferation with similar effects exhibited by the H₁ agonist 71 (Figure 7) and the H_4 agonist $-H_3$ antagonist 6, indicating involvement of H1 and H4 receptors. The H4 receptor was further implicated when the significant apoptotic effect of 6 was blocked by the H₄ antagonist JNJ-7777120. The opposite outcomes of H₃ versus H₄ receptor activation were also observed in a pancreatic carcinoma cell proliferation study.²⁵⁵ While it remains unclear whether selective antagonism of the H₃ receptor can consistently produce antiproliferative effect, inhibition of human melanoma and cutaneous carcinoma cell growth by the H₃ antagonist 5 has been reported.256

As discussed in the Supporting Information, the H₃ receptor is positively coupled via $G_{\alpha i}$ and $G_{\beta \gamma i}$ to the Ras–Raf–MEK–ERK signaling cascade. Its disregulation, often due to mutations in Ras and Raf, can contribute to aberrant cell growth and proliferation and has been linked to human cancers.²⁵⁶ Research programs are currently active targeting inhibition of this pathway through small-molecule drugs.²⁵⁷ In addition, H₃ receptor positive coupling to PI3K and Akt provides a connection to a signaling pathway most commonly activated in human cancers and also targeted for inhibition in drug development.²⁵⁸ Whether molecular connections of these pathways with H₃ receptors prove therapeutically usable remains to be seen.

Gastrointestinal Disorders

While histamine H₂ receptor antagonists have been approved for human use as antacids and antiulcer agents,³ the role of H₃ receptors in gastrointestinal disorders is not as clear. In line with high histamine levels in rapidly growing tissues, such as human neoplasias discussed above, and its apparent mitogenic activity, histamine also appears to play an important role in the self-renewing rapid turnover of gastric mucosa.²⁵⁹ The H₃ agonists 1 and 20 were shown to promote proliferation of the mucus-secreting cells and protect the overall integrity of gastric epithelium in rats, and these effects were subject to reversal by the H_3 antagonists 6 and 7.^{260,261} There is also evidence that the H3 receptor may control gastrointestinal motility in mice.²⁶² However, the potential therapeutic role of the H₃ receptor in human gastrointestinal disorders is unclear, since it has not been detected in human gastrointestinal tissues. Instead, a possibility of centrally mediated H₃ effects on acid secretion, gastrointestinal motility, and epithelial cell proliferation has been invoked.²⁶³

Cerebral Ischemia

Histamine has been shown to play an important role in mitigating the effects of cerebral ischemia. Strong reduction of

blood flow to the brain, such as resulting from temporary occlusion of blood vessels during stroke, leads to delayed morphological damage and neuronal death, even if blood flow is resumed. The underlying reason for the rapid brain damage is the combination of high energy demand and relative lack of energy reserves in neurons, leading, upon quick ATP depletion in the absence of adequate blood supply, to compromised functioning of the Na⁺/K⁺-ATPase ion pumps and resulting failure to maintain neuron membrane polarization and cellular homeostasis.²⁶⁴ From the outset, cerebral ischemia is characterized by the excessive release of glutamate (attributed to the loss of Na⁺ gradient across the cell membrane and reversed operation of glutamate transporters) which further accelerates the process of energy depletion in cells by opening additional ligand-gated ion channels, such as NMDA receptorlinked Ca²⁺ channels, and imposing an additional burden on cellular ion pumps. Increased intracellular Ca²⁺ concentration also triggers processes leading to preprogrammed cell death, while increased Na⁺ concentration causes water entry into neurons leading to cerebral edema. In a bilateral carotid artery occlusion model of ischemia in gerbil preischemic icv administration of histamine was reported to have prevented development of neuronal death.²⁶⁵ This effect of exogenous histamine was blocked by H₂, but not by H₁, antagonists, thus implicating H₂ receptors in the mediation of these effects. In addition, histamine was shown to suppress ischemic release of glutamate in the rat striatum, triggered by occlusion of the middle cerebral artery, and improve the histologic outcome.²⁶⁶ In line with the findings mentioned above, this effect was mirrored by the H₂ agonist dimaprit, whereas H₁ agonists showed no effect. Treatment with the histamine biosynthesis inhibitor 22 (Figure 7) aggravated neuronal death in a gerbil model of transient global ischemia.²⁶⁷

While the full potential of H₃ antagonists in cerebral ischemia still needs to be elucidated, their effect on histamine brain levels may be hoped to provide a beneficial outcome. Possibly complicating the overall picture is the fact that the inhibitory effect on glutamate release under nonischemic conditions is typically associated with H₃ agonists rather than antagonists.¹⁸ However, the H₃ antagonist 5, when administered in combination with L-histidine, was shown to increase brain histamine concentration and prevent development of ischemiainduced brain edema in rats, contrary to the lack of effects seen with L-histidine alone.²⁶⁸ In addition, phospholipase A₂ (PLA₂), shown to be activated in cerebral ischemia and known to contribute to CNS pathologies via increased production of reactive oxygen species, is one of the downstream effectors positively coupled to H₃ receptors, as discussed in the Supporting Information. This biochemical link may provide additional stimulus to investigate H₃ antagonists for this indication. Overall, however, one needs to appreciate the significance of the challenge by considering the general lack of progress in drug development toward cerebral ischemia across different mechanistic approaches.

Tremor

Although available information is very limited, H_3 antagonists/inverse agonists have been investigated for the treatment of movement disorders, including tremor, such as essential tremor and tremor associated with PD. In a patent application directed toward use of H_3 agonists/inverse agonsits for these indications, Merck disclosed dose-dependent activity of compound **85** (Figure 14) in the harmaline-induced rat tremor



Figure 14. Miscellaneous series of H₃ antagonists/inverse agonists.

model.²⁶⁹ While mechanistic explanation of such activity has not been suggested in the application, H₃ receptors are known to modulate dopaminergic signaling (vide supra), which, if compromised, has been implicated in motor dysfunctions, including those associated with PD itself or arising as side effects of the dopaminergic treatment (e.g., dyskinesia observed with L-dopa).²⁷⁰ In that regard, indirect pharmacological approaches present an attractive alternative. H₃ antagonists, when administered alone, have been shown to increase the release of dopamine in the cortex,¹⁵³ although not in the movement-modulating striatum.²⁷¹ However, **6** was reported to improve motor coordination in the 6-hydroxydopamine (6-OHDA) lesioned rat model of PD, and H₃ antagonism was concluded to have potential for the improvement of motor complications in PD.²⁷¹ Separately, ether **85** was mentioned in the specifications of another patent application by Merck, claiming use of H₃ antagonists/inverse agonists for the treatment of stroke (see discussion above).²⁷²

Miscellaneous Series of Preclinical Compounds

In addition to those previously mentioned, companies who have also reported H₃ antagonists at earlier stages of development include Acadia,⁵² Arena (e.g., **86**, patented crystalline form, Figure 14),²⁷³ AstraZeneca (e.g., **87**),²⁷⁴ Athersys (e.g., **88**, **89**),^{275,276} Cephalon (**90**),²⁷⁷ Eli Lilly (e.g., **91**, **92**),^{278,279} Evotec AG,⁵² Hoffman-La Roche (e.g., **93**, **94**, **95**),^{280–282} Les Laboratoires Servier (e.g., **96**),²⁸³ Neurogen (e.g., **97**, **98**),^{284,285} Pfizer (e.g., **99**, single compound patent),²⁸⁶ Sanofi-Aventis (e.g., **100**),²⁸⁷ and Wyeth (e.g., **101**).^{52,288} Patent applications claim cognition, obesity, diabetes, psychotic and sleep disorders, and allergic rhinitis as potential therapeutic applications of H₃ receptor antagonists/inverse agonists.

Conclusion

It seems likely, given the current activity level of research and development, that days of commercial H_3 drugs are approaching. Whether a stand-alone treatment or a valuable add-on to an existing therapy, H₃ antagonists seem well positioned to soon share in and expand the existing drug class designation of "antihistamines". Led by the abundant preclinical data, drug companies finally seem to be in a position to probe the putative connections between the H₃ mechanism and a number of human therapeutic conditions. The pressure is clearly on clinical research to generate convincing proof-ofconcept data to support or reject those connections. Further developments in H₃ biology are to be awaited with much interest and may serve to further substantiate the existing indications and suggest novel therapeutic uses. Because of the wide spectrum of the potential indications, the need to reconcile and manage as a combination the various expected H₃ therapeutic effects (most of which, adding to the level of complexity, are believed to be centrally mediated) may prove a significant challenge in itself.

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Supporting Information Available: Details of the H₃ receptor signaling pathways (summarized in Figure 4) and details of the clinical trials targeting H₃ receptor (as listed on www.clinicaltrials.gov). This material is available free of charge via the Internet at http://pubs.acs.org.

References

- (1) Arrang, J. M.; Garbarg, M.; Schwartz, J. C. Autoinhibition of brain histamine release mediated by a novel class (H₃) of histamine receptor. Nature 1983, 302, 832-837.
- Golightly, L. K.; Greos, L. S. Second-generation antihistamines: actions and efficacy in the management of allergic disorders. Drugs 2005, 65, 341-384.
- (3) Zeldis, J. B.; Friedman, L. S.; Isselbacher, K. J. Ranitidine: a new H₂ receptor antagonist. N. Engl. J. Med. 1983, 309, 1368-1373.
- (4)Huang, J. F.; Thurmond, R. L. The new biology of histamine
- receptors. *Curr. Allergy Asthma Rep.* **2008**, *8*, 21–27. Thurmond, R. L.; Gelfand, E. W.; Dunford, P. J. The role of histamine H_1 and H_4 receptors in allergic inflammation: the search for new antihistamines. Nat. Rev. Drug Discovery 2008, 7, 41-53.

- (6) Lovenberg, T. W.; Roland, B. L.; Wilson, S. J.; Jiang, X.; Pyati, J.; Huvar, A.; Jackson, M. R.; Erlander, M. G. Cloning and functional expression of the human histamine H₃ receptor. Mol. Pharmacol. 1999, 55, 1101-1107.
- (7) Arrang, J. M.; Garbarg, M.; Lancelot, J. C.; Lecomte, J. M.; Pollard, H.; Robba, M.; Schunack, W.; Schwartz, J. C. Highly potent and selective ligands for histamine H₃ receptors. Nature 1987, 327, 117-123.
- (8) Leurs, R.; Bakker, R. A.; Timmerman, H.; de Esch, I. J. P. The histamine H₃ receptor: from gene cloning to H₃ receptor drugs. Nat. Rev. Drug Discovery 2005, 4, 107-120 and references therein .
- (9) Bongers, G.; Bakker, R. A.; Leurs, R. Molecular aspects of the histamine H₃ receptor. Biochem. Pharmacol. 2007, 73, 1195-1204.
- (10) Yao, B. B.; Hutchins, C. W.; Carr, T. L.; Cassar, S.; Masters, J. N.; Bennani, Y. L.; Esbenshade, T. A.; Hancock, A. A. Molecular modeling and pharmacological analysis of species-related histamine H₃ receptor heterogeneity. Neuropharmacology 2003, 44, 773-786.
- (11) Pollard, H.; Moreau, J.; Arrang, J. M.; Schwartz, J. C. A detailed autoradiographic mapping of histamine H3 receptors in rat brain areas. Neuroscience 1993, 52, 169-189.
- (12) Pillot, C.; Heron, A.; Cochois, V.; Tardivel-Lacombe, J.; Ligneau, X.; Schwartz, J. C.; Arrang, J.-M. A detailed mapping of the histamine H(3) receptor and its gene transcripts in rat brain. Neuroscience 2002, 114, 173-193.
- (13) Schwartz, J.-C.; Arrang, J. M.; Garbarg, M.; Pollard, H.; Ruat, M. Histaminergic transmission in the mammalian brain. Physiol. Rev. 1991, 71, 1–51
- (14) Panula, P.; Yang, H. Y.; Costa, E. Histamine-containing neurons in the rat hypothalamus. Proc. Natl. Acad. Sci. U.S.A. 1984, 81, 2572-2576.
- (15) Watanabe, T.; Taguchi, Y.; Shiosaka, S.; Tanaka, J.; Kubota, H.; Terano, Y.; Tohyama, M.; Wada, H. Distribution of the histaminergic neuron system in the central nervous system of rats: a fluorescent immunohistochemical analysis with histidine decarboxvlase as a marker. Brain Res. 1984. 295, 13-25.
- (16) Haas, H. L. Electrophysiology of Histamine Receptors. In The Histamine Receptor; Schwartz, J. C., Haas, H. L., Eds.; Wiley-Liss: New York, 1992; pp 161-171.
- Brown, R. E.; Stevens, D. R.; Haas, H. L. The physiology of brain histamine. Prog. Neurobiol. 2001, 63, 637-672
- (18) Garduno-Torres, B.; Trevino, M.; Gutierrez, R.; Arias-Montano, J.-A. Presynaptic histamine H₃ receptors regulate glutamate, but not GABA release in rat thalamus. Neuropharmacology 2007, 52, 527-535.
- (19) Kraus, M. M.; Fischer, H.; Tran, M. H.; Philippu, A.; Prast, H. Modulation of acetylcholine release by histamine in the nucleus accumbens. Inflammation Res. 2001, 50 (Suppl. 2), S74-S75.
- (20) Schlicker, E.; Fink, K.; Hinterthaner, M.; Goethert, M. Inhibition of noradrenaline release in the rat brain cortex via presynaptic H3 receptors. Naunyn-Schmiedeberg's Arch. Pharmacol. 1989, 340, 633-638
- (21) Schlicker, E.; Fink, K.; Detzner, M.; Goethert, M. Histamine inhibits dopamine release in the mouse striatum via presynaptic H3 receptors. J. Neural Transm. 1993, 93, 1-10.
- (22) Dai, H.; Fu, Q.; Shen, Y.; Hu, W.; Zhang, Z.; Timmerman, H.; Leurs, R.; Chen, Z. The histamine H₃ receptor antagonist clobenpropit enhances GABA release to protect against NMDAinduced excitotoxicity through the cAMP/protein kinase A pathway in cultured cortical neurons. Eur. J. Pharmacol. 2007, 563, 117-123.
- (23) Threlfell, S.; Cragg, S. J.; Imre, K.; Turi, G. F.; Coen, C. W.; Greenfield, S. A. Histamine H₃ receptors inhibit serotonin release in substantia nigra pars reticulata. J. Neurosci. 2004, 24, 8704-8710.
- (24) Hofstra, C. L.; Desai, P. J.; Thurmond, R. L.; Fung-Leung, W.-P. Histamine H₄ receptor mediates chemotaxis and calcium mobilization of mast cells. J. Pharmacol. Exp. Ther. 2003, 305, 1212-1221.
- (25) Lippert, U.; Artuc, M.; Gruetzkau, A.; Babina, M.; Guhl, S.; Haase, I.; Blaschke, V.; Zachmann, K.; Knosalla, M.; Middel, P.; Krueger-Krasagakis, S.; Henz, B. M. Human skin mast cells express H₂ and H₄, but not H₃ receptors. J. Invest. Dermatol. 2004, 123, 116-123.
- (26) Rozniecki, J. J.; Letourneau, R.; Sugiultzoglu, M.; Spanos, C.; Gorbach, J.; Theoharides, T. C. Differential effect of histamine 3 receptor-active agents on brain, but not peritoneal, mast cell activation. J. Pharmacol. Exp. Ther. 1999, 290, 1427-1435.
- Varty, L. M.; Gustafson, E.; Laverty, M.; Hey, J. Activation of (27)histamine H3 receptors in human nasal mucosa inhibits sympathetic vasoconstriction. Eur. J. Pharmacol. 2004, 484, 83-89.

- (28) Arrang, J. M.; Garbarg, M.; Lancelot, J. C.; Lecomte, J. M.; Pollard, H.; Robba, M.; Schunack, W.; Schwartz, J. C. Highly potent and selective ligands for histamine H₃ receptors. *Nature* **1987**, *327*, 117–123.
- (29) Garbarg, M.; Arrang, J. M.; Rouleau, A.; Ligneau, X.; Dam Trung Tuong, M.; Schwartz, J. C.; Ganellin, C. R. S-[2-(4-Imidazolyl)ethyl]isothiourea, a highly specific and potent histamine H₃ receptor agonist. J. Pharmacol. Exp. Ther. **1992**, 263, 304–310.
- (30) Van der Goot, H.; Schepers, M. J. P.; Sterk, G. J.; Timmerman, H. Isothiourea analogues of histamine as potent agonists or antagonists of the histamine H₃ receptor. *Eur. J. Med. Chem.* 1992, 27, 511–517.
- (31) Ligneau, X.; Lin, J.-S.; Vanni-Mercier, G.; Jouvet, M.; Muir, J. L.; Ganellin, C. R.; Stark, H.; Elz, S.; Schunack, W.; Schwartz, J.-C. Neurochemical and behavioral effects of ciproxifan, a potent histamine H₃-receptor antagonist. *J. Pharmacol. Exp. Ther.* **1998**, 287, 658–666.
- (32) Gbahou, F.; Rouleau, A.; Morisset, S.; Parmentier, R.; Crochet, S.; Lin, J.-S.; Ligneau, X.; Tardivel-Lacombe, J.; Stark, H.; Schunack, W.; Ganellin, C. R.; Schwartz, J. C.; Arrang, J.-M. Protean agonism at histamine H₃ receptors in vitro and in vivo. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 11086–11091.
- (33) Arrang, J.-M.; Morisset, S.; Gbahou, F. Constitutive activity of the histamine H3 receptor. *Trends Pharmacol. Sci.* 2007, 28, 350– 357.
- (34) Greasley, P. J.; Clapham, J. C. Inverse agonism or neutral antagonism at G-protein coupled receptors: a medicinal chemistry challenge worth pursuing? *Eur. J. Pharmacol.* 2006, 553, 1–9.
- (35) Yates, S. L.; Tedford, C. E.; Gregory, R.; Pawlowski, G. P.; Handley, M. K.; Boyd, D. L.; Hough, L. B. Effects of selected histamine H₃ receptor antagonists on tele-methylhistamine levels in rat cerebral cortex. *Biochem. Pharmacol.* **1999**, *57*, 1059–1066.
- (36) Barnes, W.; Boyd, D.; Hough, L. Dynamics of histamine H₃ receptor antagonists on brain histamine metabolism: do all histamine H₃ receptor antagonists act at a single site? *Eur. J. Pharmacol.* 2001, *431*, 215–222.
- (37) Bongers, G.; Bakker, R. A.; Leurs, R. Molecular aspects of the histamine H₃ receptor. *Biochem. Pharmacol.* 2007, 73, 1195–1204.
- (38) There is fragmentary evidence pointing to possible existence of postsynaptic H₃ receptors. Such conclusion is arrived at when agonist stimulation of the H₃ receptor produces the same outcome as a parallel experiment, in which the targeted effector (histamine or otherwise) is exogenously administered or suppressed. See, for example, the following: Navarro, C. E.; Otoya, R.; Dononso, A. O. Stimulation of H₃ histamine receptors increases the release of prolactin in male rats. *Neuroendocrinology* **1993**, *57*, 654–657 as well as ref 78.
- (39) Huang, Z.-L.; Mochizuki, T.; Qu, W.-M.; Hong, Z.-Y.; Watanabe, T.; Urade, Y.; Hayaishi, O. Altered sleep-wake characteristics and lack of arousal response to H3 receptor antagonist in histamine H1 receptor knockout mice. *Proc. Natl. Acad. Sci. U.S.A.* 2006, 103, 4687–4692.
- (40) Cecchi, M.; Giorgetti, M.; Bacciottini, L.; Giovannini, M. G.; Blandina, P. Increase of acetylcholine release from cortex of freely moving rats by administration of histamine into the nucleus basalis magnocellularis. *Inflammation Res.* 1998, 47 (Suppl. 1), S32–S33.
- (41) Cecchi, M.; Pasani, M. B.; Bacciottini, L.; Mannaioni, P. F.; Blandina, P. Cortical acetycholine release elicited by stimulation of histamine H₁ receptors in the nucleus basalis magnocellularis: a dual probe microdialysis study in the freely moving rat. *Eur. J. Neurosci.* 2001, *13*, 68–78.
- (42) Blandina, P.; Giorgetti, M.; Bartolini, L.; Cecchi, M.; Timmerman, H.; Leurs, R.; Pepeu, G.; Giovannini, M. G. Inhibition of cortical acetylcholine release and cognitive performance by histamine H₃ receptor activation in rats. *Br. J. Pharmacol.* **1996**, *119*, 1656–1664.
- (43) Giorgetti, M.; Bacciottini, L.; Bianchi, L.; Giovannini, M. G.; Cecchi, M.; Blandina, P. GABAergic mechanism in histamine H₃ receptor inhibition of K⁺-evoked release of acetylcholine from rat cortex in vivo. *Inflammation Res.* **1997**, *46*, S3–S4.
- (44) Cangioli, I.; Baldi, E.; Mannaioni, P. F.; Bucherelli, C.; Blandina, P.; Passani, M. B. Activation of histaminergic H₃ receptors in the rat basolateral amygdala improves expression of fear memory and enhances acetylcholine release. *Eur. J. Neurosci.* 2002, *16*, 521–528.
- (45) Passani, M. B.; Cangioli, I.; Baldi, E.; Bucherelli, C.; Mannaioni, P. F.; Blandina, P. Histamine H₃ receptor-mediated impairment of contextual fear conditioning and in vivo inhibition of cholinergic transmission in the rat basolateral amygdala. *Eur. J. Neurosci.* 2001, 14, 1522–1532.

- (46) Takahashi, K.; Suwa, H.; Ishikawa, T.; Kotani, H. Targeted disruption of H₃ receptors results in changes in brain histamine tone leading to an obese phenotype. J. Clin. Invest. 2002, 110, 1791–1799.
- (47) Toyota, H; Dugovic, C.; Koehl, M.; Laposky, A. D.; Weber, C.; Ngo, K.; Wu, Y.; Lee, D. H.; Yanai, K.; Sakurai, E.; Watanabe, T.; Liu, C.; Chen, J.; Barbier, A. J.; Turek, F. W.; Fung-Leung, W.-P.; Lovenberg, T. W. Behavioral characterization of mice lacking histamine H₃ receptors. *Mol. Pharmacol.* **2002**, *62*, 389–397.
- ing histamine H₃ receptors. *Mol. Pharmacol.* 2002, *62*, 389–397.
 (48) Agnati, L. F.; Ferre, S.; Lluis, C.; Franco, R.; Fuxe, K. Molecular mechanisms and therapeutic implications of intramembrane receptor/receptor interactions among heptahelical receptors with examples from the striatopallidal GABA neurons. *Pharmacol. Rev.* 2003, *55*, 509–550.
- (49) Ferrada, C.; Ferre, S.; Casado, V.; Cortes, A.; Justinova, Z.; Barnes, C.; Canela, E. I.; Goldberg, S. R.; Leurs, R.; Lluis, C.; Franco, R. Interactions between histamine H₃ and dopamine D₂ receptors and the implications for striatal function. *Neurophar-macology* **2008**, *55*, 190–197.
- (50) Dopamine receptor interactions with other biogenic amine GPCRs have been exploited in drug discovery. See, for example, the following: Pinna, A. Novel investigational adenosine A_{2A} receptor antagonists for Parkinson's disease. *Expert Opin. Invest. Drugs* **2009**, *18*, 1619–1631.
- (51) Celanire, S.; Wijtmans, M.; Talaga, P.; Leurs, R.; de Esch, I. J. P. Histamine H3 receptor antagonists reach out for the clinic. *Drug Discovery Today* 2005, 10, 1613–1627 and references therein.
- (52) Investigational Drugs Database (IDdb), Thomson Pharma Partnering. http://partnering.thomson-pharma.com (accessed Aug 2009).
- (53) Stocking, E. M. Histamine H3 antagonists as wake-promoting and pro-cognitive agents. *Curr. Top. Med. Chem.* 2008, 8, 988– 1002 and references therein.
- (54) Stark, H. Histamine H3-receptor agonists and antagonists: chemical, pharmacological, and clinical aspects. *Chem. Mol. Aspects Drug Des. Action* 2008, 199, 199–214 and references therein.
- (55) Letavic, M. A.; Barbier, A. J.; Dvorak, C. A.; Carruthers, N. I. Recent medicinal chemistry of the histamine H3 receptor. *Prog. Med. Chem.* 2006, 44, 181–206.
- (56) Vohora, D. Histamine-selective H3 receptor ligands and cognitive functions: an overview. *IDrugs* 2004, 7, 667–673.
- (57) Cowart, M.; Altenbach, R.; Black, L.; Faghih, R.; Zhao, C.; Hancock, A. A. Medicinal chemistry and biological properties of non-imidazole histamine H3 antagonists. *Mini-Rev. Med. Chem.* 2004, 4, 979–992.
- (58) Wijtmans, M.; Leurs, R.; de Esch, I. Histamine H3 receptor ligands break ground in a remarkable plethora of therapeutic areas. *Expert Opin. Invest. Drugs* **2007**, *16*, 967–985.
- (59) Ramesh, V.; Thakkar, M. M.; Strecker, R. E.; Basheer, R.; McCarley, R. W. Wakefulness-inducing effects of histamine in the basal forebrain of freely moving rats. *Behav. Brain Res.* 2004, *152*, 271–278.
- (60) White, J. M.; Rumbold, G. R. Behavioural effects of histamine and its antagonists: a review. *Psychopharmacology* (*Berlin*) **1988**, 95, 1–14.
- (61) Welch, M. J.; Meltzer, E. O.; Simons, F. E. H₁-Antihistamines and the central nervous system. *Clin. Allergy Immunol.* 2002, *17*, 337–388.
- (62) Barbier, A. J.; Bradbury, M. J. Histaminergic control of sleepwake cycles: recent therapeutic advances for sleep and wake disorders. CNS Neurol. Disord.: Drug Targets 2007, 6, 31–43.
- (63) Parmentier, R.; Anaclet, C.; Guhennec, C.; Brousseau, E.; Bricout, D.; Giboulot, T.; Bozyczko-Coyne, D.; Spiegel, K.; Ohtsu, H.; Williams, M.; Lin, J. S. The brain H3-receptor as a novel therapeutic target for vigilance and sleep-wake disorders. *Biochem. Pharmacol.* 2007, *73*, 1157–1171.
- (64) Monti, J. M.; Jantos, H.; Ponzoni, A.; Monti, D. Sleep and waking during acute histamine H3 agonist BP 2.94 or H3 antagonist carboperamide (MR 16155) administration in rats. *Neuropsychopharmacology* **1996**, *15*, 31–35.
- (65) McLeod, R. L.; Aslanian, R.; Del Prado, M.; Duffy, R.; Egan, R. W.; Kreutner, W.; McQuade, R.; Hey, J. A. Sch 50971, an orally active histamine H3 receptor agonist, inhibits central neurogenic vascular inflammation and produces sedation in the guinea pig. J. Pharmacol. Exp. Ther. **1998**, 287, 43–50.
- (66) Lamberty, Y.; Margineanu, D. G.; Dassesse, D.; Klitgaard, H. H3 agonist immepip markedly reduces cortical histamine release, but only weakly promotes sleep in the rat. *Pharmacol. Res.* 2003, 48, 193–198.
- (67) Herjavecz, I.; Hirschberg, A. Histamine in Allergy. In *Histamine: Biology and Medical Aspects*; Falus, A., Ed.; Spring Med: Budapest, Hungary, 2004; pp 140–158.

- (68) Gelfand, E. W. Role of histamine in the pathophysiology of asthma: immunomodulatory and anti-inflammatory activities of H1-receptor antagonists. *Am. J. Med.* **2002**, *113*, 2S–7S.
- (69) Gross, P.; Teasdale, G. M.; Angerson, W. J.; Harper, A. M. Characteristics of increased cerebrovascular permeability resulting from carotid arterial infusion of histamine. *Prog. Migraine Res.* **1981**, *1*, 43–50.
- (70) Bell, J. K.; McQueen, D. S.; Rees, J. L. Involvement of histamine H₄ and H₁ receptors in scratching induced by histamine receptor agonists in BalbC mice. *Br. J. Pharmacol.* **2004**, *142*, 374–380.
- (71) Hossen, M. A.; Inoue, T.; Shinmei, Y.; Fujii, Y.; Watanabe, T.; Kamei, C. Role of substance P on histamine H3 antagonistinduced scratching behavior in mice. J. Pharmacol. Sci. 2006, 100, 297–302.
- (72) Yokota, E.; Kuyama, S.; Ogawa, M.; Kamei, C. Substance P is involved in the effect of histamine H3 receptor agonist, Sch 50971 on nasal allergic symptoms in mice. *Int. Immunopharmacol.* 2008, *8*, 1083–1088.
- (73) Ohkubo, T.; Shibata, M.; Inoue, M.; Kaya, H.; Takahashi, H. Regulation of substance P release mediated via prejunctional histamine H₃ receptors. *Eur. J. Pharmacol.* **1995**, *273*, 83–88.
- (74) Burgaud, J. L.; Javellaud, J.; Oudart, N. Bronchodilator action of an agonist for histamine H₃-receptors in guinea pig perfused bronchioles and lung parenchymal strips. *Lung* **1992**, *170*, 95–108.
- (75) Burgaud, J. L.; Oudart, N. Effect of an histaminergic H3 agonist on the non-adrenergic non-cholinergic contraction in guinea-pig perfused bronchioles. J. Pharm. Pharmacol. 1993, 45, 955–958.
- (76) Burgaud, J. L.; Oudart, N. Bronchodilation of guinea-pig perfused bronchioles induced by the H₃-receptor for histamine: role of epithelium. *Br. J. Pharmacol.* **1993**, *109*, 960–966.
- (77) Increased production of arachidonic acid and prostaglandin PGE₂ was effected by the H₃ agonist imetit through the MAPK-PLA₂-COX-PGE₂ signaling pathway discussed in the Supporting Information. See the following: Levi, R.; Seyedi, N.; Schaefer, U.; Estephan, R.; Mackins, C. J.; Tyler, E.; Silver, R. B. Histamine H₃ receptor signaling in cardiac sympathetic nerves: identification of a novel MAPK-PLA₂-COX-PGE₂-EP₃R pathway. *Biochem. Pharmacol.* 2007, 73, 1146–1156.
- (78) Delaunois, A.; Gustin, P.; Garbarg, M.; Ansay, M. Modulation of acetylcholine, capsaicin and substance P effects by histamine H₃ receptors in isolated perfused rabbit lungs. *Eur. J. Pharmacol.* **1995**, 277, 243–250.
- (79) Rouleau, A.; Garbarg, M.; Ligneau, X.; Mantion, C.; Lavie, P.; Advenier, C.; Lecomte, J.-M.; Krause, M.; Stark, H.; Schunack, W.; Schwartz, J.-C. Bioavailability and antinociceptive and antiinflammatory properties of BP 2-94, a histamine H3 receptor agonist prodrug. J. Pharmacol. Exp. Ther. 1997, 281, 1085– 1094.
- (80) O'Connor, B. J.; Lecomte, J. M.; Barnes, P. J. Effect of an inhaled histamine H3-receptor agonist on airway responses to sodium metabisulphite in asthma. *Br. J. Clin. Pharmacol.* **1993**, *35*, 55–57.
- (81) Millan-Guerrero, R. O.; Pineda-Lucatero, A. G.; Hernandez-Benjamin, T.; Tene, C. E.; Pacheco, M. F. *N*-Alpha-methylhistamine safety and efficacy in migraine prophylaxis: phase I and phase II studies. *Headache* 2003, 43, 389–394.
- (82) Millan-Guerrero, R. O.; Isais-Millan, R.; Benjamin, T.-H.; Tene, C. E. N-Alpha-methyl histamine safety and efficacy in migraine prophylaxis: phase III study. *Can. J. Neurol. Sci.* 2006, *33*, 195–199.
- (83) Mobarakeh, J. I.; Sakurada, S.; Katsuyama, S.; Kutsuwa, M.; Kuramasu, A.; Lin, Z. Y.; Watanabe, T.; Hashimoto, Y.; Watanabe, T.; Yanai, K. Role of histamine H1 receptor in pain perception: a study of the receptor gene knockout mice. *Eur. J. Pharmacol.* 2000, 391, 81–89.
- (84) Yoshida, A.; Mobarakeh, J. I.; Sakurai, E.; Sakurada, S.; Orito, T.; Kuramasu, A.; Kato, M.; Yanai, K. Intrathecally-administered histamine facilitates nociception through tachykinin NK1 and histamine H1 receptors: a study in histidine decarboxylase gene knockout mice. *Eur. J. Pharmacol.* 2005, *522*, 55–62.
- (85) Cannon, K. E.; Nalwalk, J. W.; Stadel, R.; Ge, P.; Lawson, D.; Silos-Santiago, I.; Hough, L. B. Activation of spinal histamine H3 receptors inhibits mechanical nociception. *Eur. J. Pharmacol.* 2003, 470, 139–147.
- (86) Cannon, K. E.; Leurs, R.; Hough, L. B. Activation of peripheral and spinal histamine H₃ receptors inhibits formalin-induced inflammation and nociception, respectively. *Pharmacol., Biochem. Behav.* 2007, 88, 122–129.
- (87) Malmberg, A. P.; Lamberti, C.; Ghelardini, C.; Giotti, A.; Bartolini, A. Role of histamine in rodent antinociception. *Br. J. Pharmacol.* **1994**, *111*, 1269–1279.
- (88) Malmberg, A. P.; Lamberti, C.; Ipponi, A.; Hanninen, J.; Gheraldini, C.; Bartolini, A. Effects of two histamine-*N*-methyltransferase inhibitors, SKF 91488 and BW 301 U, in rodent

antinociception. Naunyn-Schmiedeberg's Arch. Pharmacol. 1997, 355, 354–360.

- (89) Kennedy, J. D. Neuropathic pain: molecular complexity underlies continuing unmet medical need. J. Med. Chem. 2007, 50, 2547– 2556.
- (90) Levi, R.; Smith, N. Histamine H₃-receptors: a new frontier in myocardial ischemia. *J. Pharmacol. Exp. Ther.* 2000, *292*, 825–830.
 (91) Imamura, M.; Poli, E.; Omoniyi, A. T.; Levi, R. Unmasking of
- (91) Imamura, M.; Poli, E.; Omoniyi, A. T.; Levi, R. Unmasking of activated histamine H₃-receptors in myocardial ischemia: their role as regulators of exocytotic norepinephrine release. *J. Pharmacol. Exp. Ther.* **1994**, 271, 1259–1266.
- (92) Imamura, M.; Lander, H. M.; Levi, R. Activation of histamine H₃-receptors inhibits carrier-mediated norepinephrine release during protracted myocardial ischemia. Comparison with adenosine A₁-receptors and α₂-adrenoceptors. *Circ. Res.* **1996**, *78*, 475–481.
- (93) Fryer, R. M.; Reinhart, G. A.; Esbenshade, T. A. Histamine in cardiac sympathetic ganglia: a novel neurotransmitter? *Mol. Interventions* **2006**, *6*, 14–19.
- (94) Kumar, K. V.; Krishna, D. R.; Palit, G. Histaminergic H1 receptors mediate L-histidine-induced anxiety in elevated plusmaze test in mice. *Behav. Pharmacol.* 2007, *18*, 213–217.
- (95) Esbenshade, T. A.; Browman, K. E.; Bitner, R. S.; Strakhova, M.; Cowart, M. D.; Brioni, J. D. The histamine H3 receptor: an attractive target for the treatment of cognitive disorders. *Br. J. Pharmacol.* **2008**, *154*, 1166–1181.
- (96) Sander, K.; Kottke, T.; Stark, H. Histamine H₃ receptor antagonists go to clinics. *Biol. Pharm. Bull.* 2008, *31*, 2163–2181.
- (97) Hudkins, R. L.; Raddatz, R. Recent advances in drug discovery of histamine H3 antagonists. Annu. Rep. Med. Chem. 2007, 42, 49–62.
- (98) Esbenshade, T. A.; Fox, G. B.; Cowart, M. D. Histamine H3 receptor antagonists: preclinical promise for treating obesity and cognitive disorders. *Mol. Interventions* **2006**, *6*, 77–88.
- (99) Witkin, J. M.; Nelson, D. L. Selective histamine H3 receptor antagonists for treatment of cognitive deficiencies and other disorders of the central nervous system. *Pharmacol.Ther.* 2004, 103, 1–20 and references therein.
- (100) Vohora, D.; Pal, S. N.; Pillai, K. K. Histaminergic H3-receptors as modulators of CNS function. *Indian J. Pharmacol.* 2001, 33, 17–28.
- (101) Hancock, A. A. The challenge of drug discovery of a GPCR target: analysis of preclinical pharmacology of histamine H3 antagonists/inverse agonists. *Biochem. Pharmacol.* 2006, 71, 1103– 1113.
- (102) Petroianu, G.; Arafat, K.; Sasse, B. C.; Stark, H. Multiple enzyme inhibitions by histamine H3 receptor antagonists as potential procognitive agents. *Pharmazie* 2006, 61, 179–182.
- (103) Apelt, J.; Ligneau, X.; Pertz, H. H.; Arrang, J.-M.; Ganellin, C. R.; Schwartz, J.-C.; Schunack, W.; Stark, H. Development of a new class of nonimidazole histamine H3 receptor ligands with combined inhibitory histamine *N*-methyltransferase activity. *J. Med. Chem.* **2002**, *45*, 1128–1141.
- (104) De Esch, I.; Mills, J.; Perkins, T.; Romeo, G.; Hoffmann, M.; Wieland, K.; Leurs, R.; Menge, W.; Nederkoorn, P.; Dean, P.; Timmerman, H. Development of a pharmacophore model for histamine H₃ receptor antagonists, using the newly developed molecular modeling program SLATE. J. Med. Chem. 2001, 44, 1666–1674.
- (105) Uveges, A. J.; Kowal, D.; Zhang, Y.; Spangler, T. B.; Dunlop, J.; Semus, S.; Jones, P. G. The role of transmembrane helix 5 in agonist binding to the human H3 receptor. J. Pharmacol. Exp. Ther. 2002, 301, 451–458.
- (106) Schlegel, B.; Stark, H.; Sippl, W.; Höltje, H.-D. Model of a specific human histamine H₃ receptor (hH₃R) binding pocket suitable for virtual drug design. *Inflammation Res.* 2005, 54 (Suppl. 1), S50–S51.
- (107) Arrang, J.-M. Histamine and schizophrenia. Int. Rev. Neurobiol. 2007, 78, 247–287.
- (108) H₁ receptors are found at high concentrations in the frontal and temporal cortex, hippocampus, and thalamus regions of the human brain. See, for example, the following: Yanai, K.; Watanabe, T.; Yokoyama, H.; Hatazawa, J.; Iwata, R.; Ishiwata, K.; Meguro, K.; Itoh, M.; Takahashi, T.; Ido, T.; Matsuzawa, T. Mapping of histamine H1 receptors in the human brain using [¹¹C]pyrilamine and positron emission tomography. J. Neurochem. **1992**, 59, 128–136.
- (109) Alvarez, E. O. The role of histamine on cognition. *Behav. Brain Res.* 2009, 199, 183–189 and references therein.
- (110) Philippu, A.; Prast, H. Importance of histamine in modulatory processes, locomotion and memory. *Behav. Brain Res.* 2001, 124, 151–159 and references therein.
- (111) Yanai, K.; Dai, H.; Sakurai, E.; Watanabe, T. The roles of histamine H1 receptors on cognition. *Inflammation Res.* 2008, 57 (Suppl. 1), S39–S40.

- (112) Zlomuzica, A.; Ruocco, L. A.; Sadile, A. G.; Huston, J. P.; Dere, E. Histamine H1 receptor knockout mice exhibit impaired spatial memory in the eight-arm radial maze. *Br. J. Pharmacol.* 2009, 157, 86–91.
- (113) Terry, A. V. Role of the central cholinergic system in the therapeutics of schizophrenia. *Curr. Neuropharmacol.* 2008, 6, 286–292.
- (114) Laruelle, M.; Frankle, W. G.; Narendran, R.; Kegeles, L. S.; Abi-Dargham, A. Mechanism of action of antipsychotic drugs: from dopamine D2 receptor antagonism to glutamate NMDA facilitation. *Clin. Ther.* **2005**, *27* (Suppl. 1), S16–S24.
- (115) Prince, J. Catecholamine dysfunction in attention-deficit/hyperactivity disorder. J. Clin. Psychopharmacol. 2008, 28 (Suppl. 2), S39–S45 and references therein.
- (116) Biederman, J.; Spencer, T. Attention-deficit/hyperactivity disorder (ADHD) as a noradrenergic disorder. *Biol. Psychiatry* 1999, 46, 1234–1242.
- (117) Horner, W. E.; Johnson, D. E.; Schmidt, A. E.; Rollema, H. Methylphenidate and atomoxetine increase histamine release in rat prefrontal cortex. *Eur. J. Pharmacol.* **2007**, *558*, 96–97.
- (118) Raminder, K. Approved and investigational uses of modafinil: an evidence-based review. *Drugs* **2008**, *68*, 1803–1839.
- (119) Minzenberg, M. J.; Carter, C. S. Modafinil: a review of neurochemical actions and effects on cognition. *Neuropsychopharmacology* **2008**, *33*, 1477–1502.
- (120) Farooqui, A. A.; Horrocks, L. A. Phospholipase A₂-generated lipid mediators in the brain: the good, the bad and the ugly. *Neuroscientist* 2006, 12, 245–260.
- (121) Tedford, C. E.; Yates, S. L.; Pawlowski, G. P.; Nalwalk, J. W.; Hough, L. B.; Amin, K. M.; Phillips, J. G.; Durant, G. J.; Frederickson, R. C. A. Pharmacological characterization of GT-2016, a non-thiourea-containing histamine H3 receptor antagonist: in vitro and in vivo studies. J. Pharmacol. Exp. Ther. 1995, 275, 598–604.
- (122) Giovannini, M. G.; Bartolini, L.; Bacciottini, L.; Greco, L.; Blandina, P. Effects of histamine H3 receptor agonists and antagonists on cognitive performance and scopolamine-induced amnesia. *Behav. Brain Res.* **1999**, *104*, 147–155.
- (123) Komater, V. A.; Buckley, M. J.; Browman, K. E.; Pan, J. B.; Hancock, A. A.; Decker, M. W.; Fox, G. B. Effects of histamine H3 receptor antagonists in two models of spatial learning. *Behav. Brain Res.* 2005, 159, 295–300.
- (124) Komater, V. A.; Browman, K. E.; Curzon, P.; Hancock, A. A.; Decker, M. W.; Fox, G. B. H3 receptor blockade by thioperamide enhances cognition in rats without inducing locomotor sensitization. *Psychopharmacology* **2003**, *167*, 363–372.
- (125) Blandina, P.; Efoudebe, M.; Cenni, G.; Mannaioni, P.; Passani, M. B. Acetylcholine, histamine, and cognition: two sides of the same coin. *Learn. Mem.* **2004**, *11*, 1–8 and references therein.
- (126) Giovannini, M. G.; Efoudebe, M.; Passani, M. B.; Baldi, E.; Bucherelli, C.; Giachi, F.; Corradetti, R.; Blandina, P. Improvement in fear memory by histamine-elicited ERK2 activation in hippocampal CA3 cells. *J. Neurosci.* 2003, *23*, 9016–9023.
- (127) Akhtar, M.; Devi, P. U.; Ali, A.; Pillai, K. K.; Vohora, D. Antipsychotic-like profile of thioperamide, a selective H3-receptor antagonist in mice. *Fundam. Clin. Pharmacol.* 2006, 20, 373–378.
- (128) Jia, F.; Kato, M.; Dai, H.; Xu, A.; Okuda, T.; Sakurai, E.; Okamura, N.; Lovenberg, T. W.; Barbier, A.; Carruthers, N. I.; Iinuma, K.; Yanai, K. Effects of histamine H3 antagonists and donepezil on learning and mnemonic deficits induced by pentylenetetrazol kindling in weanling mice. *Neuropharmacology* **2006**, *50*, 404–411.
- (129) Gemkow, M. J.; Davenport, A. J.; Harich, S.; Ellenbroek, B. A.; Cesura, A.; Hallett, D. The histamine H3 receptor as a therapeutic drug target for CNS disorders. *Drug Discovery Today* 2009, 14, 509–515.
- (130) Ali, S. M.; Tedford, C. E.; Gregory, R.; Handley, M. K.; Yates, S. L.; Hirth, W. W.; Phillips, J. G. Design, synthesis, and structure– activity relationships of acetylene-based histamine H3 receptor antagonists. J. Med. Chem. 1999, 42, 903–909.
- (131) Liu, H.; Kerdesky, F. A.; Black, L. A.; Fitzgerald, M.; Henry, R.; Esbenshade, T. A.; Hancock, A. A.; Bennani, Y. L. An efficient multigram synthesis of the potent histamine H3 antagonist GT-2331 and the reassessment of the absolute configuration. *J. Org. Chem.* 2004, 69, 192–194.
- (132) Ito, S.; Yoshimoto, R.; Miyamoto, Y.; Mitobe, Y.; Nakamura, T.; Ishihara, A.; MacNeil, D. J.; Kanatani, A.; Tokita, S. Detailed pharmacological characterization of GT-2331 for the rat histamine H3 receptor. *Eur. J. Pharmacol.* **2006**, *529*, 40–46.
- (133) Stark, H; Huls, A; Ligneau, X; Purand, K.; Pertz, H; Arrang, J. M.; Schwartz, J. C.; Schunack, W. Development of FUB 181, a selective histamine H3-receptor antagonist of high oral in vivo

potency with 4-(omega-(arylalkyloxy)alkyl)-1*H*-imidazole structure. *Arch. Pharm.* **1998**, *331*, 211–218.

- (134) Meier, G.; Ligneau, X.; Pertz, H. H.; Ganellin, C. R.; Schwartz, J.-C.; Schunack, W.; Stark, H. Piperidino-hydrocarbon compounds as novel non-imidazole histamine H3-receptor antagonists. *Bioorg. Med. Chem.* 2002, *10*, 2535–2542.
- (135) Ligneau, X.; Perrin, D.; Landais, L.; Camelin, J.-C.; Calmels, T. P. G.; Berrebi-Bertrand, I.; Lecomte, J.-M.; Parmentier, R.; Anaclet, C.; Lin, J.-S.; Bertaina-Anglade, V.; la Rochelle, C. Drieu; d'Aniello, F.; Rouleau, A.; Gbahou, F.; Arrang, J.-M.; Ganellin, C. R.; Stark, H.; Schunack, W.; Schwartz, J.-C. BF2.649 [1-{3-[3-(4-chlorophenyl)propoxy]propyl}piperidine, hydrochloride], a nonimidazole inverse agonist/antagonist at the human histamine H3 receptor: preclinical pharmacology. J. Pharmacol. Exp. Ther. 2007, 320, 365–375.
- (136) www.clinicaltrials.gov. For a detailed list of clinical trials discussed in this review see the Supporting Information.
- (137) Vasudevan, A.; Conner, S. E.; Gentles, R. G.; Faghih, R.; Liu, H.; Dwight, W.; Ireland, L.; Kang, C. H.; Esbenshade, T. A.; Bennani, Y. L.; Hancock, A. A. Synthesis and evaluation of potent pyrrolidine H3 antagonists. *Bioorg. Med. Chem. Lett.* 2002, *12*, 3055–3058.
- (138) Fox, G. B.; Pan, J. B.; Radek, R. J.; Lewis, A. M.; Bitner, R. S.; Esbenshade, T. A.; Faghih, R.; Bennani, Y. L.; Williams, M.; Yao, B. B.; Decker, M. W.; Hancock, A. A. Two novel and selective nonimidazole H3 receptor antagonists A-304121 and A-317920: II. In vivo behavioral and neurophysiological characterization. J. Pharmacol. Exp. Ther. 2003, 305, 897–908.
- (139) Faghih, R.; Dwight, W.; Black, L.; Liu, H.; Gentles, R.; Phelan, K.; Esbenshade, T. A.; Ireland, L.; Miller, T. R.; Kang, C.-H.; Krueger, K. M.; Fox, G. B.; Hancock, A. A.; Bennani, Y. L. Structure-activity relationships of non-imidazole H3 receptor ligands. Part 2: Binding preference for D-amino acids motifs. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 2035–2037.
- (140) Curtis, M. P.; Dwight, W.; Pratt, J.; Cowart, M.; Esbenshade, T. A.; Krueger, K. M.; Fox, G. B.; Pan, J. B.; Pagano, T. G.; Hancock, A. A.; Faghih, R.; Bennani, Y. L. D-Amino acid homopiperazine amides: discovery of A-320436, a potent and selective non-imidazole histamine H3-receptor antagonist. *Arch. Pharm.* 2004, 337, 219–229.
- (141) Esbenshade, T. A.; Fox, G. B.; Krueger, K. M.; Baranowski, J. L.; Miller, T. R.; Kang, C. H.; Denny, L. I.; Witte, D. G.; Yao, B. B.; Pan, J. B.; Faghih, R.; Bennani, Y. L.; Williams, M.; Hancock, A. A. Pharmacological and behavioral properties of A-349821, a selective and potent human histamine H3 receptor antagonist. *Biochem. Pharmacol.* 2004, 68, 933–945.
- (142) Cowart, M.; Faghih, R.; Curtis, M. P.; Gfesser, G. A.; Bennani, Y. L.; Black, L. A.; Pan, L.; Marsh, K. C.; Sullivan, J. P.; Esbenshade, T. A.; Fox, G. B.; Hancock, A. A. 4-(2-[2-(2(R)-Methylpyrrolidin-1-yl)ethyl]benzofuran-5-yl)benzonitrile and related 2-aminoethylbenzofuran H3 receptor antagonists potently enhance cognition and attention. J. Med. Chem. 2005, 48, 38–55.
- (143) Esbenshade, T. A.; Fox, G. B.; Krueger, K. M.; Miller, T. R.; Kang, C. H.; Denny, L. I.; Witte, D. G.; Yao, B. B.; Pan, L.; Wetter, J.; Marsh, K.; Bennani, Y. L.; Cowart, M. D.; Sullivan, J. P.; Hancock, A. A. Pharmacological properties of ABT-239 [4-(2-{2-[(2R)-2-methylpyrrolidinyl]ethyl}-benzofuran-5-yl]benzonitrile]: I. Potent and selective histamine H3 receptor antagonist with drug-like properties. J. Pharmacol. Exp. Ther. 2005, 313, 165–175.
- (144) Fox, G. B.; Esbenshade, T. A.; Pan, J. B.; Radek, R. J.; Krueger, K. M.; Yao, B. B.; Browman, K. E.; Buckley, M. J.; Ballard, M. E.; Komater, V. A.; Miner, H.; Zhang, M.; Faghih, R.; Rueter, L. E.; Bitner, R. S.; Drescher, K. U.; Wetter, J.; Marsh, K.; Lemaire, M.; Porsolt, R. D.; Bennani, Y. L.; Sullivan, J. P.; Cowart, M. D.; Decker, M. W.; Hancock, A. A. Pharmacological properties of ABT-239 [4-(2-{2-[(2R)-2-methylpyrrolidinyl]ethyl}benzofuran-5-yl)benzonitrile]: II. Neurophysiological characterization and broad preclinical efficacy in cognition and schizophrenia of a potent and selective histamine H3 receptor antagonist. J. Pharmacol. Exp. Ther. 2005, 313, 176–190.
- (145) Ku, Y.-Y.; Pu, Y.-M.; Grieme, T.; Sharma, P.; Bhatia, A. V.; Cowart, M. An efficient and convergent synthesis of the potent and selective H3 antagonist ABT-239. *Tetrahedron* 2006, 62, 4584–4589.
- (146) Cowart, M.; Gfesser, G. A.; Browman, K. E.; Faghih, R.; Miller, T. R.; Milicic, I.; Baranowski, J. L.; Krueger, K. M.; Witte, D. G.; Molesky, A. L.; Komater, V. A.; Buckley, M. J.; Diaz, G. J.; Gagne, G. D.; Zhou, D.; Deng, X.; Pan, L.; Roberts, E. M.; Diehl, M. S.; Wetter, J. M.; Marsh, K. C.; Fox, G. B.; Brioni, J. D.; Esbenshade, T. A.; Hancock, A. A. Novel heterocyclic-substituted benzofuran histamine H3 receptor antagonists: in vitro

properties, drug-likeness, and behavioral activity. *Biochem. Pharmacol.* 2007, 73, 1243–1255.

- (147) Gfesser, G. A.; Faghih, R.; Bennani, Y. L.; Curtis, M. P.; Esbenshade, T. A.; Hancock, A. A.; Cowart, M. D. Structure– activity relationships of arylbenzofuran H3 receptor antagonists. *Bioorg. Med. Chem. Lett.* 2005, *15*, 2559–2563.
- (148) Altenbach, R. J.; Liu, H.; Banfor, P. N.; Browman, K. E.; Fox, G. B.; Fryer, R. M.; Komater, V. A.; Krueger, K. M.; Marsh, K.; Miller, T. R.; Pan, J. B.; Pan, L.; Sun, M.; Thiffault, C.; Wetter, J.; Zhao, C.; Zhou, D.; Esbenshade, T. A.; Hancock, A. A.; Cowart, M. D. Synthesis, potency, and in vivo profiles of quinoline containing histamine H3 receptor inverse agonists. *J. Med. Chem.* **2007**, *50*, 5439–5448.
- (149) Black, L. A.; Nersesian, D. L.; Sharma, P.; Ku, Y.-Y.; Bennani, Y. L.; Marsh, K. C.; Miller, T. R.; Esbenshade, T. A.; Hancock, A. A.; Cowart, M. 4-[6-(2-Aminoethyl)naphthalen-2-yl]benzonitriles are potent histamine H3 receptor antagonists with high CNS penetration. *Bioorg. Med. Chem. Lett.* 2007, *17*, 1443–1446.
- (150) Black, L. A.; Liu, H.; Diaz, G. J.; Fox, G. B.; Browman, K. E.; Wetter, J.; Marsh, K. C.; Miller, T. R.; Esbenshade, T. A.; Brioni, J.; Cowart, M. D. Minimization of potential hERG liability in histamine H3 receptor antagonists. *Inflammation Res.* 2008, *57*, S45–S46.
- (151) Pu, Y.-M.; Ku, Y.-Y.; Grieme, T.; Black, L. A.; Bhatia, A. V.; Cowart, M. An expedient and multikilogram synthesis of a naphthalenoid H3 antagonist. *Org. Process Res. Dev* 2007, 11, 1004–1009.
- (152) Zhao, C.; Sun, M.; Bennani, Y. L.; Gopalakrishnan, S. M.; Witte, D. G.; Miller, T. R.; Krueger, K. M.; Browman, K. E.; Thiffault, C.; Wetter, J.; Marsh, K. C.; Hancock, A. A.; Esbenshade, T. A.; Cowart, M. D. The alkaloid conessine and analogues as potent histamine H3 receptor antagonists. *J. Med. Chem.* **2008**, *51*, 5423– 5430.
- (153) Medhurst, A. D.; Atkins, A. R.; Beresford, I. J.; Brackenborough, K.; Briggs, M. A.; Calver, A. R.; Cilia, J.; Cluderay, J. E.; Crook, B.; Davis, J. B.; Davis, R. K.; Davis, R. P.; Dawson, L. A.; Foley, A. G.; Gartlon, J.; Gonzalez, M. I.; Heslop, T.; Hirst, W. D.; Jennings, C.; Jones, D. N. C.; Lacroix, L. P.; Martyn, A.; Ociepka, S.; Ray, A.; Regan, C. M.; Roberts, J. C.; Schogger, J.; Southam, E.; Stean, T. O.; Trail, B. K.; Upton, N.; Wadsworth, G.; Wald, J. A.; White, T.; Witherington, J.; Woolley, M. L.; Worby, A.; Wilson, D. M. GSK189254, a novel H3 receptor antagonist that binds to histamine H3 receptors in Alzheimer's disease brain and improves cognitive performance in preclinical models. *J. Pharmacol. Exp. Ther.* 2007, *321*, 1032–1045.
- (154) Medhurst, A. D.; Briggs, M. A.; Bruton, G.; Calver, A. R.; Chessell, I.; Crook, B.; Davis, J. B.; Davis, R. P.; Foley, A. G.; Heslop, T.; Hirst, W. D.; Medhurst, S. J.; Ociepka, S.; Ray, A.; Regan, C. M.; Sargent, B.; Schogger, J.; Stean, T. O.; Trail, B. K.; Upton, N.; White, T.; Orlek, B.; Wilson, D. M. Structurally novel histamine H3 receptor antagonists GSK207040 and GSK334429 improve scopolamine-induced memory impairment and capsaicin-induced secondary allodynia in rats. *Biochem. Pharmacol.* 2007, 73, 1182–1194.
- (155) Southam, E.; Cilia, J.; Gartlon, J. E.; Woolley, M. L.; Lacroix, L. P.; Jennings, C. A.; Cluderay, J. E.; Reavill, C.; Rourke, C.; Wilson, D. M.; Dawson, L. A.; Medhurst, A. D.; Jones, D. Preclinical investigations into the antipsychotic potential of the novel histamine H3 receptor antagonist GSK207040. *Psychopharmacology* **2009**, 201, 483–494.
- (156) Nagase, T.; Mizutani, T.; Ishikawa, S.; Sekino, E. O.; Sasaki, T.; Fujimura, T.; Ito, S.; Mitobe, Y.; Miyamoto, Y.; Yoshimoto, R.; Tanaka, T.; Ishihara, A.; Takenaga, N.; Tokita, S.; Fukami, T.; Sato, N. Synthesis, structure–activity relationships, and biological profiles of a quinazolinone class of histamine H3 receptor inverse agonists. J. Med. Chem. 2008, 51, 4780–4789.
- (157) Nagase, T.; Mizutani, T.; Sekino, E.; Ishikawa, S.; Ito, S.; Mitobe, Y.; Miyamoto, Y.; Yoshimoto, R.; Tanaka, T.; Ishihara, A.; Takenaga, N.; Tokita, S.; Sato, N. Synthesis and evaluation of structurally constrained quinazolinone derivatives as potent and selective histamine H3 receptor inverse agonists. J. Med. Chem. 2008, 51, 6889–6901.
- (158) Nagase, T.; Sato, N.; Kii, S.; Sato, K.; Tsuritani, T.; Sawada, N. Process for Production of 4(3*H*)-Quinazolinone Derivative. PCT Int. Appl. WO 2006132424, 2006.
- (159) Mizutani, T.; Nagase, T.; Ito, S.; Miyamoto, Y.; Tanaka, T.; Takenaga, N.; Tokita, S.; Sato, N. Development of novel 2-[4-(aminoalkoxy)phenyl]-4(3H)-quinazolinone derivatives as potent and selective histamine H3 receptor inverse agonists. *Bioorg. Med. Chem. Lett.* 2008, 18, 6041–6045.
- (160) Wager, T. T. Physicochemical Properties Approach to the Identification of a Histamine H3 Receptor Antagonist for the

Treatment of ADHD. *Abstracts of Papers*, 237th National Meeting of the American Chemical Society, Salt Lake City, UT, Mar 22–26, 2009; American Chemical Society: Washington, DC, 2009.

- (161) Wager, T. T.; Chandrasekaran, R. Y.; Butler, T. W. Preparation of Pyrrolidinylmethylphenylcyclobutanecarboxamides as Histamine H3 Receptor Antagonists. PCT Int. Appl. WO 2007049123, 2007.
- (162) Bembenek, S. D.; Keith, J. M.; Letavic, M. A.; Apodaca, R. D.; Barbier, A. J.; Dvorak, L.; Aluisio, L.; Miller, K. L.; Lovenberg, T. W.; Carruthers, N. I. Lead identification of acetylcholinesterase inhibitors-histamine H3 receptor antagonists from molecular modeling. *Bioorg. Med. Chem.* **2008**, *16*, 2968–2973.
- (163) Morini, G.; Comini, M.; Rivara, M.; Rivara, S.; Bordi, F.; Plazzi, P. V.; Flammini, L.; Saccani, F.; Bertoni, S.; Ballabeni, V.; Barocelli, E.; Mor, M. Synthesis and structure-activity relationships for biphenyl H3 receptor antagonists with moderate anti-cholinesterase activity. *Bioorg. Med. Chem.* 2008, *16*, 9911– 9924.
- (164) Hey, J. A.; Aslanian, R. G. Use of Dual H3 /M2 Antagonists in the Treatment of Cognition Deficit Disorders. PCT Int. Appl.WO 2002072093, 2002.
- (165) Jones, B. E. From waking to sleeping: neuronal and chemical substrates. *Trends Pharmacol. Sci.* 2005, *26*, 578–586.
 (166) Krueger, J. M.; Rector, D. M.; Roy, S.; Van Dongen, H. P. A.;
- (166) Krueger, J. M.; Rector, D. M.; Roy, S.; Van Dongen, H. P. A.; Belenky, G.; Panksepp, J. Sleep as a fundamental property of neuronal assemblies. *Nat. Rev. Neurosci.* **2008**, *9*, 910–919.
 (167) Soya, A.; Song, Y. H.; Kodama, T.; Honda, Y.; Fujiki, N.;
- (167) Soya, A.; Song, Y. H.; Kodama, T.; Honda, Y.; Fujiki, N.; Nishino, S. CSF histamine levels in rats reflect the central histamine neurotransmission. *Neurosci. Lett.* **2008**, *430*, 224–229 and references therein.
- (168) Mignot, E. Narcolepsy: Pharmacology, Pathophysiology and Genetics. In *Principles and Practice of Sleep Medicine*; Kryger, M. H., Roth, T., Dement, W. C., Eds.; Elsevier Saunders: Philadelphia, PA, 2005; pp 761–779.
- (169) Passani, M. B.; Lin, J.-S.; Hancock, A.; Crochet, S.; Blandina, P. The histamine H3 receptor as a novel therapeutic target for cognitive and sleep disorders. *Trends Pharmacol. Sci.* 2004, 25, 618–625.
- (170) Bonaventure, P.; Letavic, M.; Dugovic, C.; Wilson, S.; Aluisio, L.; Pudiak, C.; Lord, B.; Mazur, C.; Kamme, F.; Nishino, S.; Carruthers, N.; Lovenberg, T. Histamine H3 receptor antagonists: from target identification to drug leads. *Biochem. Pharmacol.* **2007**, *73*, 1084–1096.
- (171) Lin, J.-S.; Dauvilliers, Y.; Arnulf, I.; Bastuji, H.; Anaclet, C.; Parmentier, R.; Kocher, L.; Yanagisawa, M.; Lehert, P.; Ligneau, X.; Perrin, D.; Robert, P.; Roux, M.; Lecomte, J.-M.; Schwartz, J.-C. An inverse agonist of the histamine H3 receptor improves wakefulness in narcolepsy: Studies in orexin-/- mice and patients. *Neurobiol. Dis.* 2008, *30*, 74–83.
- (172) Raga, M. M.; Sallares, J.; Guerrero, M.; Guglietta, A. Preparation of 1-[3-[3-(4-Chlorophenyl)propoxy]propyl]piperidine Monohydrochloride as a Histamine H3 Receptor Ligand. PCT Int. Appl. WO 2006084833, 2006.
- (173) Guo, R. X.; Anaclet, C.; Roberts, J. C.; Parmentier, R.; Zhang, M.; Guidon, G.; Buda, C.; Sastre, J. P.; Feng, J. Q.; Franco, P.; Brown, S. H.; Upton, N.; Medhurst, A. D.; Lin, J. S. Differential effects of acute and repeat dosing with the H3 antagonist GSK189254 on the sleep-wake cycle and narcoleptic episodes in Ox-/- mice. Br. J. Pharmacol. 2009, 157, 104–117.
- (174) Le, S.; Gruner, J. A.; Mathiasen, J. R.; Marino, M. J.; Schaffhauser, H. Correlation between ex vivo receptor occupancy and wake-promoting activity of selective H3 receptor antagonists. *J. Pharmacol. Exp. Ther.* **2008**, *325*, 902–909.
- (175) Shah, C.; McAtee, L.; Breitenbucher, J. G.; Rudolph, D.; Li, X.; Lovenberg, T. W.; Mazur, C.; Wilson, S. J.; Carruthers, N. I. Novel human histamine H3 receptor antagonists. *Bioorg. Med. Chem. Lett.* 2002, *12*, 3309–3312.
- (176) Apodaca, R.; Dvorak, C. A.; Xiao, W.; Barbier, A. J.; Boggs, J. D.; Wilson, S. J.; Lovenberg, T. W.; Carruthers, N. I. A new class of diamine-based human histamine H3 receptor antagonists: 4-(aminoalkoxy)benzylamines. J. Med. Chem. 2003, 46, 3938– 3944.
- (177) Barbier, A. J.; Berridge, C.; Dugovic, C.; Laposky, A. D.; Wilson, S. J.; Boggs, J.; Aluisio, L.; Lord, B.; Mazur, C.; Pudiak, C. M.; Langlois, X.; Xiao, W.; Apodaca, R.; Carruthers, N. I.; Lovenberg, T. W. Acute wake-promoting actions of JNJ-5207852, a novel, diamine-based H3 antagonist. *Br. J. Pharmacol.* **2004**, *143*, 649–661.
- (178) Dvorak, C. A.; Apodaca, R.; Barbier, A. J.; Berridge, C. W.; Wilson, S. J.; Boggs, J. D.; Xiao, W.; Lovenberg, T. W.; Carruthers, N. I. 4-Phenoxypiperidines: potent, conformationally restricted, non-imidazole histamine H3 antagonists. J. Med. Chem. 2005, 48, 2229–2238.

- (179) Fujiki, N.; Yoshino, F.; Lovenberg, T. W.; Nishino, S. Wake promoting effects of non-imidazoline histamine H3 antagonist in orexin/ataxin-3 narcoleptic mice. *Sleep (Sleep Res. Soc. Meet.)* 2006, 29, No. A230.
- (180) Galici, R.; Boggs, J. D.; Aluisio, L.; Fraser, I. C.; Bonaventure, P.; Lord, B.; Lovenberg, T. W. JNJ-10181457, a selective nonimidazole histamine H3 receptor antagonist, normalizes acetylcholine neurotransmission and has efficacy in translational rat models of cognition. *Neuropharmacology* 2009, *56*, 1131–1137.
- (181) Mani, N. S.; Jablonowski, J. A.; Jones, T. K. A scalable synthesis of a histamine H3 receptor antagonist. J. Org. Chem. 2004, 69, 8115–8117.
- (182) Nagase, T.; Sato, N.; Kanatani, A.; Tokita, S. Preparation of a 4(3*H*)-Quinazolinone Compound as Histamine H3 Receptor Antagonist or Inverse Agonist. PCT Int. Appl. WO 2006085692, 2006.
- (183) Dunlop, B. W.; Crits-Christoph, P.; Evans, D. L.; Hirschowitz, J.; Solvason, H. B.; Rickels, K.; Garlow, S. J.; Gallop, R. J.; Ninan, P. T. Coadministration of modafinil and a selective serotonin reuptake inhibitor from the initiation of treatment of major depressive disorder with fatigue and sleepiness: a double-blind, placebo-controlled study. *J. Clin. Psychopharmacol.* 2007, *27*, 614–619.
- (184) Imeri, L.; Opp, M. R. How (and why) the immune system makes us sleep. *Nat. Rev. Neurosci.* 2009, 10, 199–210.
- (185) Barbier, A. J.; Aluisio, L.; Lord, B.; Qu, Y.; Wilson, S. J.; Boggs, J. D.; Bonaventure, P.; Miller, K.; Fraser, I.; Dvorak, L.; Pudiak, C.; Dugovic, C.; Shelton, J.; Mazur, C.; Letavic, M. A.; Carruthers, N. I.; Lovenberg, T. W. Pharmacological characterization of JNJ-28583867, a histamine H3 receptor antagonist and serotonin reuptake inhibitor. *Eur. J. Pharmacol.* 2007, *576*, 43–54.
- (186) Letavic, M. A.; Keith, J. M.; Jablonowski, J. A.; Stocking, E. M.; Gomez, L. A.; Ly, K. S.; Miller, J. M.; Barbier, A. J.; Bonaventure, P.; Boggs, J. D.; Wilson, S. J.; Miller, K. L.; Lord, B.; McAllister, H. M.; Tognarelli, D. J.; Wu, J.; Abad, M. C.; Schubert, C.; Lovenberg, T. W.; Carruthers, N. I. Novel tetrahydroisoquinolines are histamine H3 antagonists and serotonin reuptake inhibitors. *Bioorg. Med. Chem. Lett.* 2007, *17*, 1047– 1051.
- (187) Smith, F. M.; Haskelberg, H.; Tracey, D. J.; Moalem-Taylor, G. Role of histamine H3 and H4 receptors in mechanical hyperalgesia following peripheral nerve injury. *NeuroImmunoModulation* 2007, 14, 317–325.
- (188) Lim, H.; van Rijn, R.; Ling, P.; Bakker, R.; Thurmond, R.; Leurs, R. Evaluation of histamine H1-, H2-, and H3-receptor ligands at the human histamine H4 receptor: identification of 4-methylhistamine as the first potent and selective H4 receptor agonist. J. Pharmacol. Exp. Ther. 2005, 314, 1310–1321.
- (189) Owen, S. M.; Sturman, G.; Freeman, P. Modulation of morphineinduced antinociception in mice by histamine H3-receptor ligands. *Agents Actions* 1994, 41, C62–C63.
- (190) Farzin, D.; Asghari, L.; Nowrouzi, M. Rodent antinociception following acute treatment with different histamine receptor agonists and antagonists. *Pharmacol., Biochem. Behav.* 2002, 72, 751–760.
- (191) Huang, L.; Adachi, N.; Nagaro, T.; Liu, K.; Arai, T. Histaminergic involvement in neuropathic pain produced by partial ligation of the sciatic nerve in rats. *Reg. Anesth. Pain Med.* 2007, *32*, 124–129.
- (192) Medhurst, S. J.; Collins, S. D.; Billinton, A.; Bingham, S.; Dalziel, R. G.; Brass, A.; Roberts, J. C.; Medhurst, A. D.; Chessell, I. P. Novel histamine H3 receptor antagonists GSK189254 and GSK334429 are efficacious in surgically-induced and virally-induced rat models of neuropathic pain. *Pain* 2008, *138*, 61–69.
- (193) Sindrup, S. H.; Otto, M.; Finnerup, N. B.; Jensen, T. S. Antidepressants in the treatment of neuropathic pain. *Basic Clin. Pharmacol. Toxicol.* **2005**, *96*, 399–409.
- (194) Yokoyama, H; Iinuma, K; Yanai, K; Watanabe, T; Sakurai, E; Onodera, K. Proconvulsant effect of ketotifen, a histamine H1 antagonist, confirmed by the use of D-chlorpheniramine with monitoring electroencephalography. *Methods Find. Exp. Clin. Pharmacol.* **1993**, *15*, 183–188.
- (195) Vohora, D.; Pal, S. N.; Pillai, K. K. Histamine as an anticonvulsant inhibitory neurotransmitter. *Curr. Neuropharmacol.* 2004, 2, 419–425.
- (196) Crowley, V.; Yeo, G.; O'Rahilly, S. Obesity therapy: altering the energy intake-and-expenditure balance sheet. *Nat. Rev. Drug Discovery* 2002, 1, 276–286.
- (197) Clineschmidt, B. V.; Lotti, V. J. Histamine: intraventricular injection suppresses ingestive behavior of the cat. Arch. Int. Pharmacodyn. Ther. 1973, 206, 288–298.

- (198) Itowi, N.; Nagai, K.; Nakagawa, H.; Watanabe, T.; Wada, H. Changes in the feeding behavior of rats elicited by histamine infussion. *Physiol. Behav.* **1998**, *44*, 221–226.
- (199) Lecklin, A.; Etu-Seppala, P.; Stark, H.; Tuomisto, L. Effects of intracerebroventricularly infused histamine and selective H1, H2 and H3 agonists on food and water intake and urine flow in Wistar rats. *Brain Res.* **1998**, *793*, 279–288.
- (200) Sheiner, J. B.; Morris, P.; Anderson, G. H. Food intake suppression by histidine. *Pharmacol.*, *Biochem. Behav.* 1985, 23, 721–726.
- (201) Lecklin, A.; Tuomisto, L.; McDonald, E. Metoprine, an inhibitor of histamine N-methyl transferase but not cathecol-O-methyl transferase, suppresses feeding in sated and in food deprived rats. *Methods Find. Exp. Clin. Pharmacol.* **1995**, *17*, 47–52.
- (202) Orthen-Gambill, N.; Salomon, M. FMH-induced decrease in central histamine levels produced increased feeding and body weight in rats. *Physiol. Behav.* **1992**, *51*, 891–893.
- (203) Sakata, T.; Ookuma, K.; Fukagawa, K.; Fujimoto, K.; Yoshimatsu, H.; Shiraishi, T.; Wada, H. Blockade of the histamine H1 receptor in the rat ventromedial hypothalamus and feeding elicitation. *Brain Res.* 1988, 441, 403–407.
- (204) Baptista, T.; Zarate, J.; Joober, R.; Colasante, C.; Beaulieu, S.; Paez, X.; Hernandez, L. Drug induced weigh gain, an impediment to successful pharmacotherapy: focus on antipsychotics. *Curr. Drug Targets* 2004, 5, 279–299.
- (205) Ookuma, K.; Sakata, T.; Fujimoto, K. Evidence for feeding elicited through antihistaminergic effects of tricyclic antidepressants in the rat hypothalamus. *Psychopharmacology* **1990**, *101*, 481–485.
- (206) Richelson, E.; Souder, T. Binding of antipsychotic drugs to human brain receptors. Focus on newer generation compounds. *Life Sci.* 2000, *68*, 29–39.
- (207) Horak, F.; Subner, U. P. Comparative tolerability of second generation antihistamines. *Drug Saf.* **1999**, *20*, 385–401.
- (208) Masaki, T.; Yoshimatsu, H.; Chiba, S.; Watanabe, T.; Sakata, T. Targeted disruption of histamine H1-receptor attenuates regulatory effects of leptin on feeding, adiposity, and UCP family in mice. *Diabetes* 2001, 50, 385–391.
- (209) Jorgensen, E. A.; Vogelsang, T. A.; Knigge, U. Increased susceptibility to diet-induced obesity in histamine-deficient mice. *Neuroendocrinology* 2006, *83*, 289–294.
- (210) See for example: Pan, J.; Yao, B.; Miller, T.; Kroeger, P.; Bennani, Y.; Komater, V.; Esbenshade, T.; Hancock, A.; Decker, M.; Fox, G. Evidence for tolerance following repeated dosing in rats with ciproxifan, but not with A-304121. *Life Sci.* 2006, 79, 1366– 1379.
- (211) Leurs, R.; Smit, M.; Bast, A.; Timmerman, H. Homologous histamine H1 receptor desensitization results in reduction of H1 receptor agonist efficacy. *Eur. J. Pharmacol.* **1991**, *196*, 319–322.
- (212) Hancock, A. A.; Brune, M. E. Assessment of pharmacology and potential antiobesity properties of H3 receptor antagonists/ inverse agonists. *Expert Opin. Drugs* 2005, 14, 223–241.
- (213) Sakata, T.; Kazuyoshi, O.; Fujimoto, K.; Fukagawa, K.; Yoshimatsu, H. Histaminergic control of energy balance in rats. *Brain Res. Bull.* **1991**, *27*, 371–375.
- (214) Yoshimoto, R.; Miyamoto, Y.; Shimamura, K.; Ishihara, A.; Takahashi, K.; Kotani, H.; Chen, A. S.; Chen, H. Y.; MacNeil, D. J.; Kanatani, A.; Tokita, S. Therapeutic potential of histamine H3 receptor agonist for the treatment of obesity and diabetes mellits. *Proc. Natl. Acad. Sci. U.S.A.* 2006, *103*, 13866–13871.
- (215) Tokita, S. Histamine H3 Receptor as a Potential Target for Metabolic Disorders. Presented at the 3rd Modern Drug Discovery and Development Summit, San Francisco, CA, November 28–30, 2007.
- (216) Puigserver, P.; Wu, Z.; Park, C. W.; Graves, R.; Wright, M.; Spiegelman, B. M. A cold-inducible coactivator of nuclear receptors linked to adaptive thermogenesis. *Cell* **1998**, *92*, 829–839.
- (217) Weyer, C.; Gautier, J. F.; Danforth, E. Development of β3-adrenoceptor agonists for the treatment of obesity and diabetes—an update. *Diabetes Metab.* 1999, 25, 11–21.
- (218) Sandoval, D. A., Obici, S.; Seeley, R. J. Targeting the CNS to treat type 2 diabetes. *Nat. Rev. Drug Discovery* **2009**, *8*, 386–398 and references therein .
- (219) Masaki, T.; Yoshimatsu, H.; Chiba, S.; Watanabe, T.; Sakata, T. Central infusion of histamine reduces fat accumulation and up-regulates UCP family in leptin resistant obese mice. *Diabetes* 2001, 50, 376–384.
- (220) Fumihiko, H.; Ichiro, N. Diabetes Model Mouse. Jpn. Pat. Appl. JP 2004065181, 2004.
- (221) Yamamoto, Y.; Ishino, F.; Kaneko-Ishino, T.; Shiura, H.; Uchio-Yamada, K.; Matsuda, J.; Suzuki, O.; Sato, K. Type 2 diabetes mellitus in a non-obese mouse model induced by Meg1/ Grb10 overexpression. *Exp. Anim.* **2008**, *57*, 385–395.

- (222) Masaki, T.; Yoshimatsu, H. Neuronal histamine and its receptors in obesity and diabetes. Curr. Diabetes Rev. 2007, 3, 212-216.
- (223)Moniri, N. H.; Covington-Strachan, D.; Booth, R. G. Liganddirected functional heterogeneity of histamine H1 receptors: novel dual-function ligands selectively activate and block H1-mediated phospholipase C and adenylyl cyclase signaling. J. Pharmacol. Exp. Ther. 2004, 311, 274–281 and references therein.
- (224) Arrang, J. M.; Garbarg, M.; Quach, T. T. Actions of betahistine at histamine receptors in the brain. Eur. J. Pharmacol. 1985, 111, 73-84
- (225) Fossati, A.; Barone, D.; Benvenuti, C. Binding affinity profile of betahistine and its metabolites for central histamine receptors of rodents. Pharmacol. Res. 2001, 43, 389-392.
- (226) Rossi, R. Effect of the H1-histamine receptor agonist betahistine on drinking and eating behavior in pygmy goats. Physiol. Behav. 1999, 66, 517–521.
- (227) Szelag, A. Betahistine inhibits food intake in rats. Pol. J. Pharmacol. 1999, 53, 701-707.
- (228) Barak, N.; Greenway, F. L.; Fujioka, K.; Aronne, L. J.; Kushner, R. F. Effect of histaminergic manipulation on weight in obese adults: a randomized placebo controlled trial. Int. J. Obes. 2008, 32, 1559-1565.
- (229) Hancock, A. A.; Bennani, Y. L.; Bush, E. N.; Esbenshade, T. A.; Faghih, R.; Fox, G. B.; Jacobson, P.; Knourek-Segel, V.; Krueger, K. M.; Nuss, M. E.; Pan, J. B.; Shapiro, R.; Witte, D. G.; Yao, B. B. Antiobesity effects of A-331440, a novel nonimidazole histamine H3 receptor antagonist. Eur. J. Pharmacol. 2004, 487, 183-197
- (230) Hancock, A. A.; Diehl, M. S.; Fey, T. A.; Bush, E. N.; Faghih, R.; Miller, T. R.; Krueger, K. M.; Pratt, J. K.; Cowart, M. D.; Dickinson, R. W.; Shapiro, R.; Knourek-Segel, V. E.; Droz, B. A.; McDowell, C. A.; Krishna, G.; Brune, M. E.; Esbenshade, T. A.; Jacobson, P. B. Antiobesity evaluation of histamine H3 receptor antagonist analogs of A-331440 with improved safety (231) Zaragoza, F.; Stephensen, H.; Knudsen, S. M.; Pridal, L.; Wulff,
- B. S.; Rimvall, K. 1-Alkyl-4-acylpiperazines as a new class of imidazole-free histamine H3 antagonists. J. Med. Chem. 2004, 47, 2833-2838
- (232) Malmlof, K.; Zaragoza, F.; Golozoubova, V.; Refsgaard, H. H. F.; Cremers, T.; Raun, K.; Wulff, B. S.; Johansen, P. B.; Westerink, B.; Rimvall, K. Influence of a selective histamine H3 receptor antagonist on hypothalamic neural activity, food intake and body weight. Int. J. Obes. 2005, 29, 1402-1412.
- (233) Peschke, B.; Bak, S.; Hohlweg, R.; Petterson, I.; Refsgaard, H. H. F.; Viuff, D.; Rimvall, K. Cinnamic amides of (S)-2-(aminomethyl)pyrrolidines are potent H3 antagonists. Bioorg. Med. Chem. Lett. 2004, 12, 2603-2616.
- (234) Malmlof, K.; Golozoubova, V.; Peschke, B.; Wulff, B. S.; Refsgaard, H. H. F.; Johansen, P. B.; Cremers, T.; Rimvall, K. Increase of neuronal histamine in obese rats is associated with decreases in body weight and plasma triglycerides. Obesity 2006, 14.2154-2162
- (235) Malmlof, K.; Hastrup, S.; Wulff, B. S.; Hansen, B. C.; Peschke, B.; Jeppesen, C. B.; Hohlweg, R.; Rimvall, K. Antagonistic targeting of the histamine H3 receptor decreases caloric intake in higher mammalian species. Biochem. Pharmacol. 2007, 73, 1237 - 1242
- (236) Holgate, S. T.; Polosa, R. Treatment strategies for allergies and asthma. Nat. Rev. Immunol. 2008, 8, 218-230.
- (237) Doyle, W. J.; Boehm, S.; Skoner, D. P. Physiologic responses to intranasal dose-response challenges with histamine, methacholine, bradykinin, and prostaglandin in adult volunteers with and without nasal allergy. J. Allergy Clin. Inmunol. 1990, 86, 924-935.
- (238) Hey, J.; Aslanian, R.; Bolster, D.; Chapman, R.; Egan, R.; Rizzo, C.; Shih, N.-Y.; Fernandez, Z.; McLeod, R.; West, R.; Kreutner, W. Studies on the pharmacology of the novel histamine H3 receptor agonist Sch 50971. Arzneim-Forsch/Drug Res. 1998, 48, 881-888.
- (239) McLeod, R. L.; Mingo, G. G.; Herczku, C. Combined histamine H1 and H3 receptor blockade produces nasal decongestion in an experimental model of nasal congestion. Am. J. Rhinol. 1999, 13, 391-399
- (240) Hancock, A. P.; Hodgson, S. T.; Vinader Brugarolas, M.-V.; Washington, M. L. 1-{4-[(1-Cyclobutyl-4-piperidinyl)oxy]phenyl]-4-{[4-(methylsulfonyl)phenyl]carbonyl Piperazine as Histamine H3 Antagonist. PCT Int. Appl. WO 2006090142, 2006.
- (241) Gore, P. M.; Hancock, A. P.; Hodgson, S. T.; Kindon, L. J.; Procopiou, P. A. 2-Substituted-4-benzylphthalazinone Derivatives as Histamine H1 and H3 Antagonists. PCT Int. Appl. WO 2007122156, 2007.

- (242) Hodgson, S.; Procopiou, P.; Vinader, B. Preparation of 3-[4-[[4-[4-[[3-(3,3-Dimethyl-1-piperidinyl)propyl]oxy]phenyl]-1-piperidinyl]carbonyl]-1-naphthalenyl]propanoates as Histamine H1 and H3 Antagonists for the Treatment of Inflammatory and/or Allergic Disorders. PCT Int. Appl. WO 2007071691, 2007
- (243) McLeod, R. L.; Rizzo, C. A.; West, R. E.; Aslanian, R. G.; McCormick, K. D.; Bryant, M.; Hsieh, Y.; Korfmacher, W.; Mingo, G. G.; Varty, L.; Williams, S. M.; Shih, N.-Y.; Egan, R. W.; Hey, J. A. Pharmacological characterization of the novel histamine H3 receptor antagonist N-(3,5-dichlorophenyl)-N'-[[4-(1H-imidazol-4-ylmethyl)phenyl]-methyl]urea (Sch 79687). *J. Pharmacol. Exp. Ther.* **2003**, *305*, 1037–1044. (244) Aslanian, R.; Mutahi, M.; Shih, N.-Y.; Piwinski, J.; West, R.;
- Williams, S.; She, S.; Wu, R.-L.; Hey, J. Identification of a dual H1/H3 receptor ligand based on the H1 antagonist chlorpheniramine. Bioorg. Med. Chem. Lett. 2003, 13, 1959-1961.
- (245) Arrang, J. M.; Garbarg, M.; Quach, T. T. An agent for the amelioration of vertigo in Meniere's syndrome. Betahistine hydrochloride (Serc). JAMA, J. Am. Med. Assoc. 1968, 203, 1122.
- (246) James, A. L.; Burton, M. J. Betahistine for Meniere's disease or syndrome. Cochrane Database Syst. Rev. 2001, 1, No. CD001873.
- (247)Anonymous. Treatment of Meniere's disease. Br. Med. J. 1970, 614-616.
- (248) Tomita, M.; Gotoh, F.; Sato, T.; Amano, T.; Tanahashi, N.; Tanaka, K.; Yamamoto, M. Comparative responses of the carotid and vertebral arterial systems of rhesus monkeys to betahistine. Stroke 1978, 9, 382-387.
- (249) Smith, K.; Meyer, M. Distribution of cardiac output in dogs during intravenous infusion of betahistine. Stroke 1976, 7, 257–260.
- (250) Panula, P.; Lintunen, M.; Karlstedt, K. Histamine in brain
- (251) Fantai, F., Bani, C., Kanaka, K., K Capaccioli, S.; Masini, E. The role of cyclooxygenase-2 in mediating the effects of histamine on cell proliferation and vascular endothelial growth factor production in colorectal cancer. Clin. Cancer Res. 2005, 11, 6807-6815.
- (252) Medina, V.; Cricco, G.; Nuñez, M.; Martin, G.; Mohamad, N.; Correa-Fiz, F.; Sanchez-Jimenez, F.; Bergoc, R.; Rivera, E. Histamine-mediated signaling processes in human malignant mammary cells. Cancer Biol. Ther. 2006, 5, 1462-1471.
- (253) Sieja, K.; Stanosz, S.; Von Mach-Szczypinski, J.; Olewniezak, S.; Stanosz, M. Concentration of histamine in serum and tissues of the primary ductal breast cancer in women. Breast 2005, 14, 236-241 and references therein
- (254) Medina, V.; Croci, M.; Crescenti, E.; Mohamad, N.; Sanchez-Jimenez, F.; Massari, N.; Nuñez, M.; Cricco, G.; Martin, G.; Bergoc, R.; Rivera, E. The role of histamine in human mammary carcinogenesis: H3 and H4 receptors as potential therapeutic targets for breast cancer treatment. Cancer Biol. Ther. 2008, 7, 28-35.
- (255) Cricco, G.; Mohamad, N.; Sambuco, L.; Genre, F.; Croci, M.; Gutierrez, A.; Medina, V.; Bergoc, R.; Rivera, E.; Martin, G. Histamine regulates pancreatic carcinoma cell growth through H₃ and H4 receptors. Inflammation Res. 2008, 57 (Suppl. 1), S23-S24.
- (256)Szelag, A. Role of histamine H3-receptors in the proliferation of neoplastic cells in vitro. Med. Sci. Monitor 1998, 4, 747-755.
- McCubrey, J.; Milella, M.; Tafuri, A.; Martelli, A.; Lunghi, P. (2.57)Bonati, A.; Cervello, M.; Lee, J.; Steelman, L. Targeting the Raf/ MEK/ERK pathway with small-molecule inhibitors. Curr. Opin. Invest. Drugs 2008, 9, 614-630.
- (258) Liu, P.; Cheng, H.; Roberts, T.; Zhao, J. Targeting the phosphoinositide 3-kinase pathway in cancer. Nat. Rev. Drug Discovery 2009, 8, 627-644.
- (259) Coruzzi, G.; Poli, E.; Morini, G.; Bertaccini, G. The Histamine H3 Receptor: Pharmacotherapy Targets in Gastrointestinal Disorders. In Drug Development: Molecular Targets for GI Diseases; Gaginella, T. S., Guglietta, A., Eds.; Humana Press: Totowa, NJ, 1999; pp 239-267.
- (260) Morini, G.; Grandi, D.; Schunack, W. Ligands for histamine H3 receptors modulate cell proliferation and migration in rat oxyntic mucosa. Br. J. Pharmacol. 2002, 137, 237-244.
- (261) Grandi, D.; Schunack, W.; Morini, G. Epithelial cell proliferation is promoted by the histamine H3 receptor agonist (\hat{R})- α -methylhistamine throughout the rat gastrointestinal tract. Eur. J. Pharmacol. 2006, 538, 141-147.
- Pozzoli, C.; Todorov, S.; Schunack, W.; Timmerman, H.; Coruzzi, (262)G.; Poli, E. Role of histamine H3 receptors in control of mouse intestinal motility in vivo and in vitro: comparison with α -2 adrenoreceptors. Dig. Dis. Sci. 2002, 47 (5), 1065-1072.
- (263) Poli, E.; Pozzoli, C.; Coruzzi, G. Role of histamine H3 receptors in the control of gastrointestinal motility. An overview. J. Physiol. 2001, 95, 67-74.

- (264) Adachi, N. Cerebral ischemia and brain histamine. *Brain Res. Rev.* **2005**, *50*, 275–286 and references therein .
- (265) Fujitani, T.; Adachi, N.; Nagaro, T.; Miyazaki, H.; Nakamura, Y.; Kataoka, K.; Arai, T. Histaminergic H₂ action protects hippocampal CA1 neurons by prolonging the onset of the anoxic depolarization in gerbils. *J. Neurochem.* **1996**, *67*, 2613–2615.
- (266) Hamami, G.; Adachi, N.; Liu, K.; Arai, T. Alleviation of ischemic neuronal damage by histamine H₂ receptor stimulation in the rat striatum. *Eur. J. Pharmacol.* 2004, *484*, 167–173.
 (267) Sugimoto, K.; Abe, K.; Lee, T. H.; Sakurai, E.; Yanai, K.;
- (267) Sugimoto, K.; Abe, K.; Lee, T. H.; Sakurai, E.; Yanai, K.; Kogure, K.; Itoyama, Y.; Watanabe, T. Histamine depletion in brain caused by treatment with (*S*)α-fluoromethylhistidine enhances ischemic damage of gerbil hippocampal CA2 neurons. *Brain Res.* **1994**, *666*, 279–283.
 (268) Irisawa, Y.; Adachi, N.; Liu, K.; Arai, T.; Nagaro, T. Alleviation
- (268) Irisawa, Y.; Adachi, N.; Liu, K.; Arai, T.; Nagaro, T. Alleviation of ischemia-induced brain edema by activation of the central histaminergic system in rats. J. Pharmacol. Sci. 2008, 108, 112–123.
- (269) Marino, M.; Seabrook, G. Treatment of Tremor or Other Movement Disorder with Histamine H3 Inverse Agonists or Histamine H3 Antagonists. PCT Int. Appl. WO 2005097111, 2005.
- (270) Kostrzewa, R. M.; Nowak, P.; Kostrzewa, J. P.; Kostrzewa, R. A.; Brus, R. Peculiarities of L-DOPA treatment of Parkinson's disease. *Amino Acids* 2005, 28, 157–164.
- (271) Nowak, P.; Bortel, A.; Dabrowska, J.; Biedka, I; Slomian, G.; Roczniak, W.; Kostrzewa, R. M.; Brus, R. Histamine H₃ receptor ligands modulate L-dopa-evoked behavioral responses and L-dopa-derived extracellular dopamine in dopamine-denervated rat striatum. *Neurotoxic. Res.* 2008, *13*, 231–240.
- (272) Seabrook, G.; Koblan, K.; Ho, T. W.-H. Treatment of Stroke with Histamine H3 Inverse Agonists or Histamine H3 Antagonists. PCT Int. Appl. WO 2006058023, 2006.
- (273) Blackburn, A.; Demattei, J.; Hart, R.; Khulman, Y.; Mesleh, M.; Smith, J. Crystalline Forms of (R)-1-{2-[4'- (3-Methoxy-propane-1-sulfonyl)-biphenyl-4-yl]-ethyl}-2-methyl-pyrrolidine, and Compositions, and Methods Related Thereto. PCT Int. Appl. WO 2008153958, 2008.
- (274) Folmer, J.; Hunt, S.; Hamley, P.; Wesolowski, S. Preparation of Piperidine Derivatives as Histamine H3 Receptor Ligands for Treatment of Depression. PCT Int. Appl. WO 2006014134, 2006.
- (275) Bennani, Y.; Anderson, J.; Wang, J.; Campbell, M. Preparation of Heterocyclic Non-Imidazole Tertiary Amines as Histamine 3 Receptor Inhibitors for the Treatment of Cognitive and Sleep Disorders, Obesity and Other CNS Disorders. PCT Int. Appl. WO 2006004937, 2006.
- (276) Bennani, Y.; Anderson, J.; Wang, J.; Campbell, M. Preparation of 4-Piperidinol Tertiary Amines as Histamine 3 Receptor Inhibitors for the Treatment of Obesity and Other CNS Disorders. PCT Int. Appl. WO 2006071750, 2006.

- (277) Hudkins, R.; Josef, A.; Tao, M. Preparation of Pyridizinones as Histamine H3 Inhibitors. PCT Int. Appl. WO 2008013838, 2008
- (278) Hipskind, P.; Takakuwa, T.; Jesudason, C.; Gadski, R.; Hornback, W.; Pickard, R.; Beavers, L. Preparation of Phenyl Pyrrolidinylmethylpyrrolidinyl Methanones as Histamine H3 Receptor Agents. PCT Int. Appl. WO 2006107661, 2006.
- (279) Jesudason, C.; Beavers, L.; Cramer, J.; Dill, J.; Finley, D.; Lindsley, C.; Stevens, F.; Gadski, R.; Oldham, S.; Pickard, R.; Siedem, C.; Sindelar, D.; Singh, A.; Watson, B.; Hipskind, P. Synthesis and SAR of novel histamine H3 receptor antagonists. *Bioorg. Med. Chem. Lett.* 2006, *16*, 3415–3418.
- (280) Roche, O.; Nettekoven, M.; Vifian, W.; Sarmiento, R. Refinement of histamine H3 ligands pharmacophore model leads to a new class of potent and selective naphthalene inverse agonists. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 4377–4379.
- (281) Roche, O.; Sarmiento, R. A new class of histamine H3 receptor antagonists derived from ligand based design. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 3670–3675.
- (282) Pierson, P.; Fettes, A.; Freichel, C.; Gatti-McArthur, S.; Hertel, C.; Huwyler, J.; Mohr, P.; Nakagawa, T.; Nettekoven, M.; Plancher, J.-M.; Raab, S.; Richter, H.; Roche, O.; Sarmiento, R.; Schmitt, M.; Schuler, F.; Takahashi, T.; Taylor, S.; Ullmer, C.; Wiegand, R. 5-Hydroxyindole-2-carboxylic acid amides: novel histamine-3 receptor inverse agonists for the treatment of obesity. J. Med. Chem. 2009, 52, 3855–3868.
- (283) Casara, P.; Chollet, A.; Dhainaut, A.; Bert, L.; Lestage, P.; Lockhart, B. New Azabicyclic, Particularly Octahydrocyclopenta[c]pyrrole, Derivatives as H3 Receptor Antagonists, Process for Their Preparation, and Pharmaceutical Compositions Containing Them. PCT Int. Appl. WO 2005089747, 2005.
- (284) Pringle, W.; Peterson, J.; Xie, L.; Ge, P.; Gao, Y.; Ochterski, J.; Lan, J. Preparation of Thiazole Amides, Imidazole Amides and Related Analogues as Histamine H3 Receptor Modulators. PCT Int. Appl. WO 2006089076, 2006.
- (285) Gao, Ŷ.; Han, B.; Xu, Y.; Caldwell, T.; Xie, L. Preparation of Piperazinyloxoalkyl Tetrahydroisoquinolines and Related Analogues as Histamine H3 Receptor Modulators. PCT Int. Appl. WO 2007106349, 2007.
- (286) Lunn, G. Tetrahydronaphthyridine Derivative, Its Preparation, and Its Therapeutic Use. PCT Int. Appl. WO 2007052124, 2007.
- (287) Diaz, J.; Escribano, B.; Martin, U. Cyclopropylmethanone Derivatives, Particularly (Cyclopropyl)(phenyl)methanone Derivatives, and Their Preparation and Therapeutic Use as H3 Histamine Receptor. PCT Int. Appl. WO 2005037810, 2005.
 (288) Cole, D. C.; Asselin, M.; Stock, J. R.; Kim, J.-I. Preparation of
- (288) Cole, D. C.; Asselin, M.; Stock, J. R.; Kim, J.-I. Preparation of N-Benzoyl- and N-Benzylpyrrolidin-3-ylamines as Histamine-3 Antagonists. PCT Int. Appl. WO 2007136668, 2007.